Identification of epulis patients with a high risk of periodontal diseases: DNA infrared spectra, expression MGMT and p53 by and apoptosis analysis

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Background

The objective of this study was to epulis apoptosis analyze, expression of MGMT and p53, DNA methylation patients with a high risk of developing bone resorption and inflammatory periodontal diseases.

Methods.

Human epulis benign were assessed for their ability to expression of MGMT, p53. 51 epulis DNA were compared by infrared spectroscopy for their ability to apoptosis induced. We conducted a comparative analysis of global expression changes in human epulis using K-S test and Student method .

Results.

MGMT and p53 was expressed in giant-cell. Expression of MGMT and p53 in giant-cell epulis. By immunohistochemistry, 97.1±0.42% (P<0.001) of giant-cells were positive for MGMT, whereas only 6.21±0.26% of giant-cells were positive for p53 (P<0.001). Induction of the enzymatic activity of MGMT was increased p53. This could be explained by the fact that MGMT is physiologically expressed by the giant-cell of epulis, p53 expression is weak or absent in giant-cell epulis and non induced apoptosis in the fibrous connective tissue.

Significant changes were observed IR absorption bands at - δsSN3 group bases. In intact periodontium in CH3 - IR bands were unchanged. Center band oscillations - δsSN3 group was at 1375 ± 1cm - 1. Percentage intensity on a scale transmission of infrared radiation was 7,18 ± 0,74%. Percentage of infrared absorption bands 1375 ± 1 cm- 1 in giant cell epulis equal to 13,24±3,7% (** P = 0.01).

Changes in IR absorption bands observed in δsCH2 group. In intact periodontal δsCH2 next strip - strip center fluctuations - 1464 cm- 1, the percentage of intensity on a scale transmission of infrared radiation - 0,24 ± 0,03%. Percentage of IR absorption bands - 1464 cm-1 in the giant cell epulis was

0,46±0,09% (* P = 0.05). Percent transmittance intensity on a scale stretching, deformation and rocking vibrations of CH3 and CH2 groups of DNA

Conclusion.

Aim of this study was to find if IR can be used in neoplasm process DNA methylation detection. Statistical analysis shows that there are differences between spectra of giant cell epulis and control group. Fitting analysis allows to follow small changes in spectra CH3 groups. Presented results prove that infrared spectroscopy could be useful tool in DNA methylation. Morphologically, apoptosis is characterized by DNA fragmentation and formation of apoptotic. DNA CH3 groups can be detected by infrared spectra leads of breaks in DNA strands. This leads to the activation of apoptosis via p53 and MGMT.

Keywords: MGMT and p53; immunohistochemistry; oral epulis.

INTRODUCTION

Gingivitis represents a spectrum of diseases whose origin is commonly attributed to the presence of bacteria, but there are other forms of gingivitis that are not primarily plaque-related. Colonization of periodontal tissues gram-positive and gram-negative periodontal bacteria including: P gingivalis, Actinomycetemcomitans, T forsythia, T denticola, B Forsythus, P Gingivalis F Nucleatum, P intermedius, P nigrescens, and P micros, S Sanguis, S oralis, S mitis, S gordonii, S intermedius, Capnocytophagia, C concius and E corrodens appears to be the primary initiator of the disease [1]. Many oral bacteria, the ability to invade periodontal tissues cells and establish an intracellular space is a critical survival mechanism. As exemplified by P Gingivalis, this initially innocuous relationship with a periodontal cell can potentially shift to a more worse [2].

Plaque bacteria evoking the chronic inflammatory response in periodontal tissues [3]. At the same time, it was reported that destructive processes responsible for periodontal tissue breakdown, leading to the clinical of periodontitis [4]. Therefore, the characteristic features of chronic periodontitis occur mainly as a result of activation of the host-derived immune and inflammatory defense mechanism.

Systemic diseases such as diabetes and leukemia can exacerbate plaque-associated gingivitis, as can endocrine changes (puberty, pregnancy), medications (nifedipine, cyclosporineand phenytoin) and malnutrition (vitamin C deficiency) [5].

Epulis - these overgrowths induced by chronic inflammatory. Kramer's investigation is a convincing periapical irritation induce periodontal disease [6]. In severe cases it's associated with loss of alveolar crest bone. However this is uncommon and it's only confined to the soft tissue. More common in female probably due to hormonal defect, most affected area is anterior to molar teeth [7]. Now epulides can be classified (according to clinical appearance, histopathologic appearance and sometimes the origin) into fibrous epulis, ossificans epulis, pyogenic granuloma (also known as, angina epulis, hemangioma), giant cell epulis (also known as osteoclastoma, giant cell reparative granuloma, peripheral giant cell granuloma or giant cell hyperplasia) and combined epulis.

Specific T- and B-cell–mediated antibacterial responses activate a network of regulatory cytokines (IFNG gene) that are produced by infiltrating T-helper type. The methylation pattern of the promoter region CpG from the IFNG gene have studed Kelma Campos et. al 2013 [8]. Bobetsis et al [9] presented partial or total methylation of the promoter region of the gene IFNG that is epigenetic alteration during inflammatory.

IFNG gene in experimentally induced gingivitis independent of promoter methylation alteration. Hypomethylation IFNG gene promoter region is related to an increase of IFNG transcription present in the chronic periodontitis biopsies. [10]

A study by Yin and Chung [11] says that bacterial infection (Campylobacter rectus) induced hypermethylation of the placental f2 promoter of mice. It is well accepted that exposure to different oral bacteria results in differential methylation profiles. Recent evidence indicates that changes related to methylation patterns can occur in periodontitis.

The hypermethylation of E-cadherin, COX-2 and PTGS-2 genes have described in gingival biopsy samples from individuals with chronic periodontitis [12, 13]. Other studies [14, 15] have suggested that the hypomethylation of IL-8 gene is present in oral epithelial cells from individuals with aggressive periodontitis and with chronic periodontitis.

In [19] there was a discuss about hypermethylation status of E- Cadherin and COX-2 genes. Wings TY Looet al [19] analysis shown correlated among the three groups with statistical significance ($p < 0.0001$). The methylation of CpG islands in E-Cadherin and COX-2 genes in periodontitis patients occurs more frequently in periodontitis patients than in thecontrol subjects, but occurs less frequently than in the breast cancer patients

Hypermethylation in the IFN-γ and IL-10 genes Michelle B. V. et al is a first time investigated in healthy and inflamed periodontal tissues [16].

In recent years, our understanding of the processes controlling the dynamics of partial methylation of IL-6 gene has been considerably investigated by Florenc Abdanur Stefani et al [17].

The overall methylation of the studied PTGS2 promoter region (-541 bp ~ -216 bp) in chronically inflamed gingival tissues has 5.06-fold higher than the methylation level exhibited in non-inflamed gingival tissues [18].

The results Naila F. P. et al indicate that individuals with chronic periodontitis, independent of smoking habit, have a higher percentage of hipomethylation of the IL8 gene than those controls in epithelial oral cells ($p=0.0001$), and expression of higher levels of interleukin-8 (IL-8) mRNA than controls in gingival cells. Naila F. P. et al has conclude that inflammation in the oral mucosa might lead to changes in the DNA methylation status of the IL8 gene in epithelial oral cells [20].

The most dangerous endogenous DNA methylating agent is S-adenosylmethionine (SAM) [21]. SAM is a coenzyme involved in more than 40 metabolic reactions transfer of methyl groups on nucleic acids [22]. On the one

hand, enzymatic DNA methylation is necessary for the regulation of gene expression, on the other hand - can lead to mutagenesis of DNA [23].

During the marginal periodontitis inflammation in the cell SAM able to create: thousand 7-Methylguanosine (m7G), hundreds - 3 methyladenine (m3A) and tens - of Methylguanosine 6^{th} (m6G) [24]. The first two adduct contribute to the formation of AP sites or blocking DNA replication.

MGMT protein acts through a self-destruction mechanism, removing abnormal adducts from the O6 position of guanine, providing protection from mutagenic agents [25]. Loss of MGMT expression has been associated with aggressive tumor behavior and progression in several types of neoplasia, including esophageal, hepatocellular, lung, gastric and breast carcinomas [26].

Many types of stress activate p53, including DNA damage, telomere attrition, oncogene activation, hypoxia and loss of normal growth and survival signals. These stress signals may be encountered by a developing tumor. Activation of p53 can induce several responses in cells, including differentiation, senescence, DNA repair and the inhibition of angiogenesis [27].

So it is necessary to understand the cellular mechanism of degeneration in periodontal tissues in various types of epulis. DNA methylation under the influence of chronic inflammatory could be key to better understand disease evolution.

In the present work, we show the expression of MGMT and p53 in epulis. There have been no reports about expression of MGMTand p53 in various types of epulis.

METHODS

The study sample consisted of periodontal and epulis tissues of patients. The subjects were divided into two equal groups:

Patient’s Group (Group I): included 51people who had clinical diagnosis of epulis. Only patients with available tissue represent a subset of the overall study groups.

Control group (Group II): included 9 patients who died in Sumy Regional Hospital Patients with various diagnoses (not atherosclerotic ones).

The infrared spectroscopy was used for gathering structural information on biological systems, but not used in periodontitis inflammation researchers. The study of DNA by infrared spectroscopy requires peeled DNA samples. The infrared specters of DNA show many characteristic: denaturation, alkylation, dehydration and conformational transition.

DNA was isolated from epulis and periodontal tissues using buffer (30 mMTrisCl; 10 mM EDTA; 1% SDS; proteinaza-K.). DNA purification was performed standard phenolic-chloroform method followed by precipitation in absolute ethanol. The resulting DNA product was triturated with and embedded in KBr tablet subsequent FTIR spectrophotometer Spectrum One (Perkin Elmer).

Hematoxylin and eosin (H&E) stains have been used for at least a century and are still essential for recognizing various tissue types and the morphologic change. Paraffin sections are immersing into three sets of xylene for 10 minutes each followed by three sets of absolute ethanol for 10 minutes and finally rinsed with tap water. Slides are placed into haematoxylin for 5 minutes and rinsed thoroughly under tap water for approximately 4–5 minutes. Excess haematoxylin is removed by adding 1% acid alcohol (1% HCl in 70%(v/v) alcohol) for 5 seconds followed by a tap water wash. The slides are rinsed in tap water before being stained in eosin (1% (w/v)) for 15 seconds with a subsequent wash in running tap water for 1–5 minutes. The slides are covered with glass cover slips.

Paraffin sections were prepared for acridine orange staining by mounting on superfrost slides, drying a hot plate, and then immersing into three sets of xylene for 2 minutes each followed by three sets of absolute ethanol for 5 minutes and finally rinsed with tap water. The aim was to remove the wax and dehydrate the sections. Slides (paraffin) were placed into acridine orange staining solution for 15 minutes, and rinsed with phosphate-buffered saline (PBS). Then the slide was soaked in 0.1% calcium chloride solution for 3 minutes and was washed with PBS

once again. Cover glass was mounted for observation under a fluorescence microscope to observe and read the result.

Immunostainings for MGMT and p53, were performed on formalin-fixed (pH 7,4), paraffin-embedded thyroid tissue sections using mouse monoclonal anti-MGMT, and anti- p53 (Thermo Fisher Scientific UK). Briefly, 4μm thick tissue sections were dewaxed in xylene and were brought to water through graded alcohols. Antigen retrieval was performed by microwaving slides in 10mM citrate buffer (pH 6.2) for 30 min at high power, according to the manufacturer's instructions. To remove the endogenous peroxidase activity, sections were then treated with freshly prepared 1.0% hydrogen peroxide in the dark for 30 min at 37°C temperature. Non-specific antibody binding was blocked using blocking serum. The sections were incubated for 30 min, at 37°C temperature, with the primaries antibodies against MGMT and p53, diluted 1:100 in phosphate buffered saline (PBS) pH 7.2, after washing 3 times with PBS. Anti-(mouse IgG)–horseradish peroxidase conjugate (1:40 000 dilution) was used for the detection of the MGMT, p53, primaries antibodies, sections were then incubated for 20 min, at 37° C temperature. The colour was developed by DAB.

Appearance of positive factors was detected semiquantitatively by counting of positive structures in visual field (+ - few, ++ - moderate, +++ - numerous, ++++ - abundance positive structures in visual field.

Results were presented as mean values (±SD). The K-S test was used in order to evaluate the normality of the data. Also, the Student method was used to perform simple comparative analysis A value of $p<0.05$ was considered significant.

RESULTS

A total of 51 cases including: 20 fibrous epulis,4 ossificans epulis, 9 pyogenic granuloma, 14 giant cell epulis and 4 combined epulis diagnosed and treated in the surgeon of maxillofacial department in Sumy Regional Hospital and, Sumy Regional Oncology Center Ukraine from 2012 to 2014 were retrospectively analysed. Epulides were classified basing on histopathological diagnosis.

The patients were divided into four age groups, ie., Group I – up to 18 years; Group II – 18 to 40 years; Group III – 41 to 60 years; and Group IV – over the age of 60

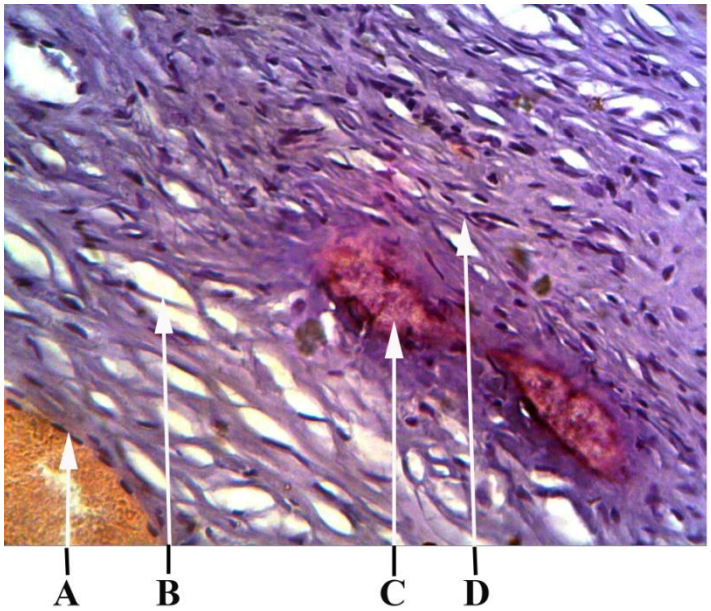

Fig.1 Fibrous epulis, Haematoxylin and Eosin (x800 magnification) A – capillaries, B – edema areas, C – resorption areas, D - connective tissue.

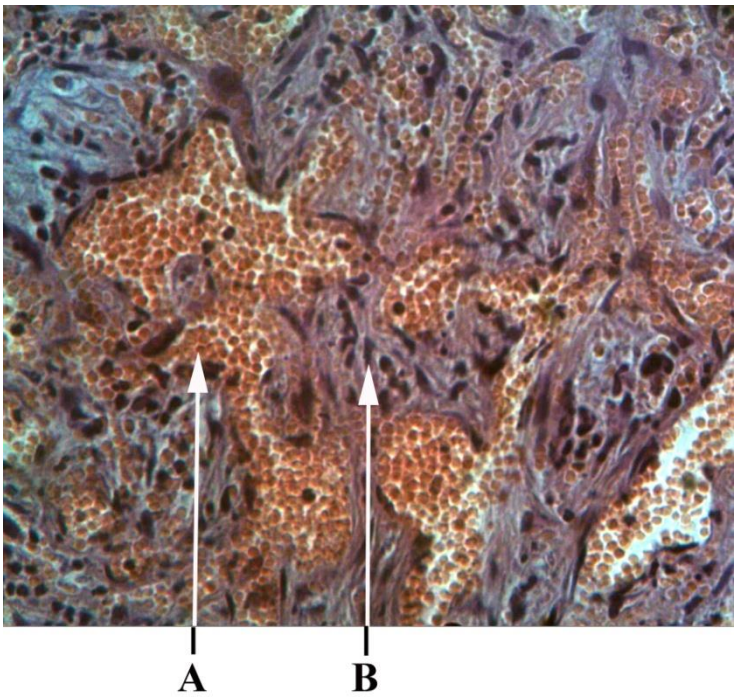

Fig.2 Pyogenic epulis, Haematoxylin and Eosin (x800 magnification) A – vascular spaces**,** B - connective tissue

The microscopic examination of the hematoxylin and eosin stained section shown, a single of tissue with epithelium and underlying fibrous connective tissue

stroma (Fig.1).The sections revealed well-circumscribed, fibroblasts (Fig.1 D), capillaries (Fig.3 A), edema areas (Fig.3 B) and occasionally there are bone resorption areas (Fig.1 C). There were also several areas characterised by haemorrhage, underlined by the presence of Fe deposits.

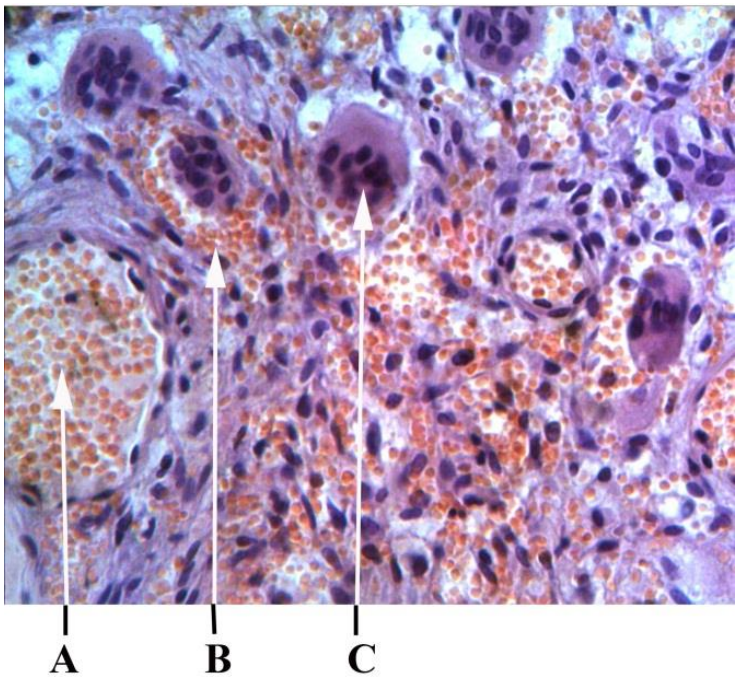

Fig 3Giant-cell epulis, Haematoxylin and Eosin (x400 magnification) A – capillaries, B - Areas of haemorrhages, C - Giant cells

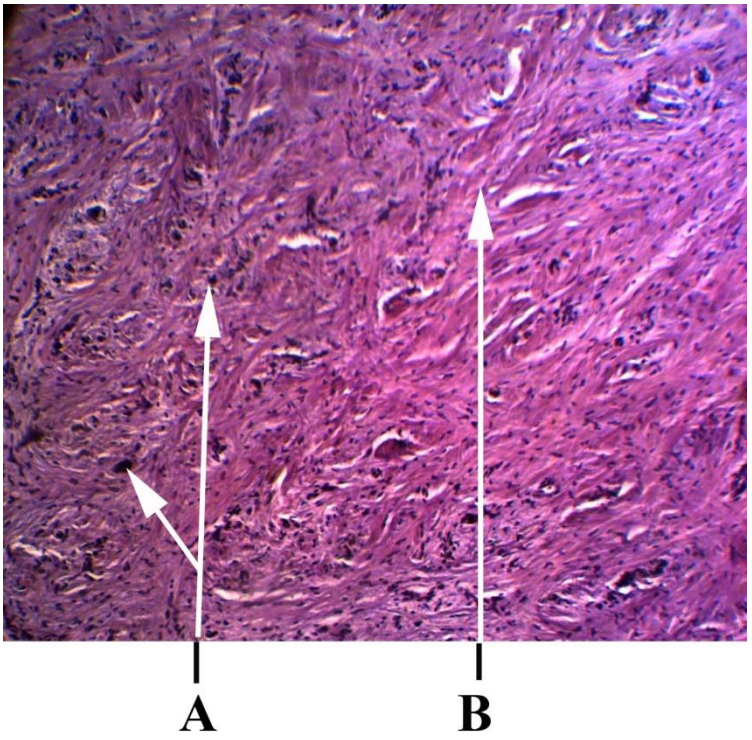

Fig.4 Ossificans epulis, Haematoxylin and Eosin (x100 magnification) A – acellular mineralization, B - fibroblasts

These microscopic features are typical for a pyogenic epulis. Hematoxylin and eosin stained pyogenic epulis sections of the specimen (Fig.2) exhibited fibrovascular connective tissue with overlying ulcerated stratified squamous epithelium with features of atrophy and proliferation at different places. The ulcerated area was covered by fibrinous exudate. The connective tissue (Fig.2 B) was fibrocellular with abundant vascularity. There were numerous endothelium lined vascular spaces (Fig.2 A), budding endothelial cells and proliferation of fibroblasts. A moderate degree of chronic inflammatory cell infiltrate, composed chiefly of lymphocytes and plasma cells, was present.

The sections revealed well-circumscribed, unencapsulated cellular mass containing oval to spindle-shaped fibroblasts, abundant multinucleated giant cells (Fig. 3 C), numerous capillaries (Fig. 3 A) and areas of haemorrhage (Fig.3 B). These giant cells were localized in the deep corion in a vascular stroma of ovoid and spindle shaped fibroblastys. The multinucleated giant cells were of variable shapes and sizes containing open-faced nuclei ranging from 10 to 25 in number conforming to the type I giant cells described in literature. There were also several areas characterised by haemorrhage, underlined by the presence of Fe deposits.

The diagnosis was ossificans epulis with spindle-shaped fibroblasts (Fig.4 B). There were no atypias or mitotic figures presence of a fusocellular tumor containing calcifications with concentric and acellular mineralization (Fig.4 A) at the center and other areas presenting recently formed osteoid with peripheral osteoblasts and signs of progressive calcifcation**.**

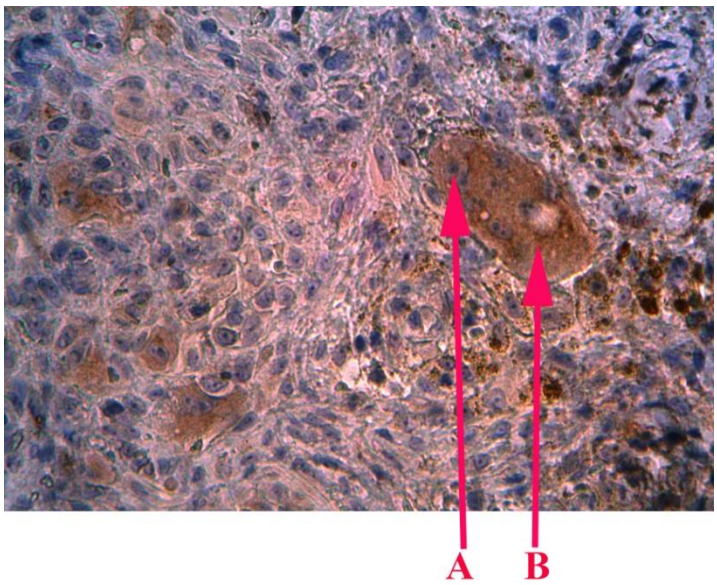

Fig 5Expression of MGMT proteins in giant-cell epulis(x800 magnification) A - Giant cells MGMT “-”nucleus, B - Giant cells MGMT “+”cytoplasm

MGMT and p53 was expressed in giant-cell. Expression of MGMT and p53 in giant-cell epulises shown in Figures 5 and 6.By immunohistochemistry, 97.1±0.42%(P<0.001) of giant-cells were positive for MGMT, whereas only 6.21±0.26% of giant-cells were positive for p53 (P<0.001). Induction of the enzymatic activity of MGMT was increased p53. This could be explained by the fact that MGMT is physiologically expressed by the giant-cell of epulis, p53 expression is weak or absent in giant-cell epulis and non induced apoptosis in the fibrous connective tissue.

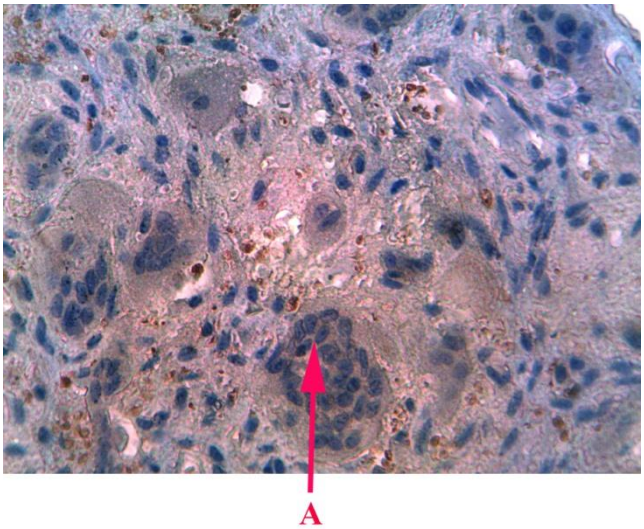

Fig6Expression of p53 proteins in giant-cell epulis, (x800 magnification) A - Giant cells p53 “-”.

P53 immunoexpression was negative in the 18 cases of fibrous epulis and positive in the case of simple dysplasia from the papilloma type, confirming the changes noted in usual staining (Fig7). In all of the fibrous epulis, MGMT was positive (98.54±10.19% positive nuclei).

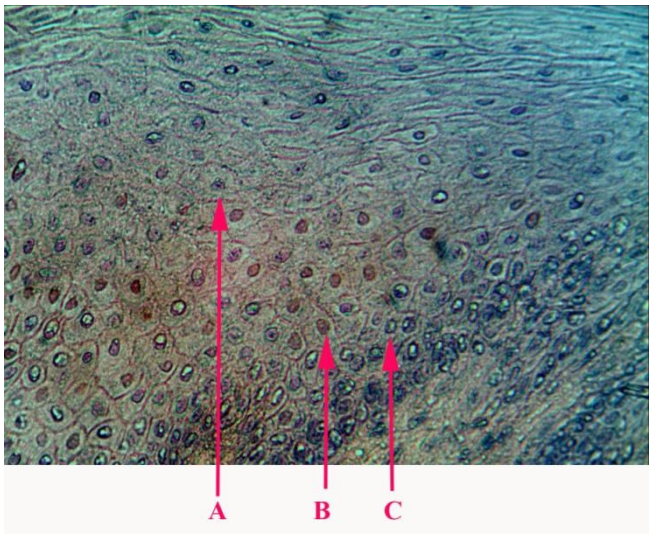

Fig7 Expression of p53 proteins indysplasia from the papilloma type(x600 magnification) A - cells p53 "-" B - cells p53 "+" C - basal cells

Positive MGMT immunostaining was present in 87.2±7.20% of ossificans epulis. Positive p53 immunostaining was found in 23.00±2.02%.ofossificans epulis.

All pyogenic epulis immunopositivity for MGMT in fibroblasts and in cellular infiltration. Interestingly, in the periphery of the lesion these ones showed a moderate positivity around the blood vessels to MGMT 24.35±4.89% related antigen, reaction not evident deeper in the fibroblasts and, in cellular infiltration.In 9 of the samples analyzed, at least one the p53 sample was positive.

Infrared spectra (IR) of the DNA bands can be roughly divides into three areas: first - 4000-2000 cm-1 - variations of bases, the second - 1700-1500 cm- 1 - DNA deoxyribose vibrations, third - 1300- 1000 cm -1 - deoxyribose vibrations and phosphate groups in the skeleton of the DNA molecule. The bands of the spectrum depending on the absorption of infrared radiation can be dividedinto: strong - ≤ 20% average - 20 % -5% and weak - 5 % ≥. Due to the complexity of the

structure of DNA arises imposition peaks of adenine, thymine, guanine, cytosine, deoxyribose and phosphorus balance in the group CH3, CH2.

The most important differentiation CH3 groups attached to abnormal bases from CH 2 groups deoxyribose. Stretching vibrations of the C-H fragments alkyl groups CH3, CH2 appear in the 3000-2840 cm- 1. In the area spectra 3000 - 2840 cm-1 arises partly overlay peaks of adenine, thymine, guanine, cytosine. For differentiation groups CH3, CH2, remember that stretching vibrations Ssp3 -H bonds are usually observed below 3000 cm-1 , while stretching vibrations links Ssp2 -N and Ssp -N lie above 3000 cm- 1 (fig 8).

Stretching vibrations of methyl groups (CH3) are observed as two bands at 2962 and 2872 cm- 1.

The first - the result of antisymmetric stretching vibrations in which the two C-H stretching of the methyl group, while the third connectivity is compressed (νas CH3).

The second band is due to symmetric stretching vibrations (νs CH3), when all three C-H bonds are stretched or compressed in phase. The presence of several methyl groups leads to an increase in the intensity of the respective bands.

Stretching vibrations of methylene groups (CH 2) are also observed in two bands (2962 and 2853 cm- 1) due to antisymmetric (νas CH2) and symmetric (νs CH2) stretching vibrations.

In the methyl groups may show two deformation vibrations: symmetric deformational vibrations (δs CH3), which is found around 1375 cm- 1 and antisymmetric deformational vibrations (δas CH3) - in the 1450 cm- 1.

The absorption at 1375 cm-1 is an important criterion (δs CH3) group. Negligible absorption band in the spectrum of DNA and characterized compression intact DNA. Methyl group has four types of deformation vibrations (scissor, fan-shaped,shuttle,spinning). The most informative is the absorption in the 1465 cm-1 due to scissor deformation vibrations (δs CH2). Therefore, CH3 and CH2 comparison groups in DNA intact periodontium and periodontal epulides of the above, we analyze in the absorption band.

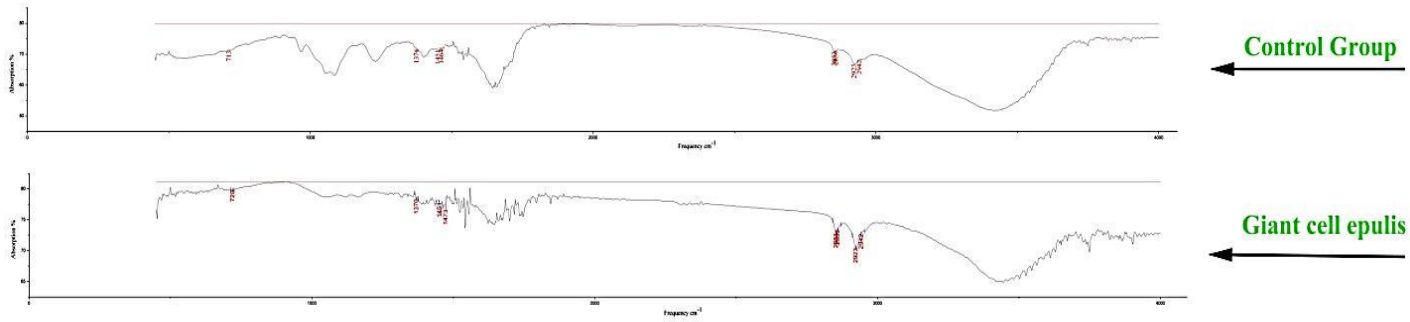

Fig 8 DNA IR specter.

Significant changes were observed IR absorption bands at - δsSN3 group bases. In intact periodontium in CH3 - IR bands were unchanged. Center band oscillations - δsSN3 group was at 1375 ± 1 cm -1. Percentage intensity on a scale transmission of infrared radiation was 7,18 ± 0,74%. Percentage of infrared absorption bands 1375 ± 1 cm- 1 in giant cell epulis equal to 13,24±3,7% (** P = 0.01).

Changes in IR absorption bands observed in δsCH2 group. In intact periodontal δsCH2 next strip - strip center fluctuations - 1464 cm-1, the percentage of intensity on a scale transmission of infrared radiation - 0,24 ± 0,03%. Percentage of IR absorption bands - 1464 cm-1 in the giant cell epulis was 0,46±0,09% (* P = 0.05). Percent transmittance intensity on a scale stretching, deformation and rocking vibrations of CH3 and CH2 groups of DNA are shown in Table № 1.

Table № 1.

Percentage intensity on a scale transmission of infrared radiation CH3 and CH2 groups of DNA.

	2953 (vasCH3)	2870 (vsCH3)	2922 (vasCH2)	2853 (vsCH2)	**1465 (δsCH2)**	1450 (δasCH3)	**1375 (δsCH3)**	724(pCH2)
	Stretchingvibrations mean ±SD				Deformationvibrations mean ±SD			Pendulumos cillation mean ±SD
Control Group**N =9**	48,05 ±4,3	0,37± 0,01	3,17± 0,16	18,97 ±3,77	0,24± 0,03	8,88± 1,36	7,18± 0,74	3,08±1,07
fibrous epulis N=20	45,5 ±3,3	0,42± 0,13	4,0±0, 6	19,7± 2,75	0,33± 0,021	10,21 ±2,62	7,94± 1,57	4,12±0,89*
ossifica nsepulis N=4	47,97 ±2,7 8	0,4±0, 06	3,4±0, 78	20,97 ±4,21 *	0,31± 0,07	9,26± 3,08	8,11± 2,42	2,89±1,00
pyogeni c granulo ma N=9	50,1 ±6,6 *	0,27± 0,05	4,2±0, 81	21,31 ±4,28	0,42± 0,081	11,29 ±2,61	10,34 ±2,67	5,1±0,72*
giant cell epulis N=14	53,9 ±3,7 1***	0,72± 0,25*	3,22± 0,92	25,94 ±3,89 *	0,46± 0,09*	15,47 ±1,16 **	13,24 ±3,7* *	7,33±0,59**
combin ed epulis N=4	49,1 ±3,9 *	0,52± 0,09*	3,0±0, 95	19,19 ±4,81	0,31± 0,1	9,21± 5,18	9,41± 4,31	4,1±0,21
*P=0.05, **P=0.01, ***P=0.001								

In most model systems, etoposide-induced apoptosis is dependent on p53 [52]. Acridine orange is a nucleic acid selective metachromatic stain useful for cell cycle determination. Acridine orange interacts with DNA and RNA by intercalation or electrostatic attraction respectively. DNA intercalated acridine orange fluoresces green (525nm); RNA electrostatically bound acridine orange fluoresces red (>630nm). It may distinguish between quiescent and activated, proliferating cells, and may also allow differential detection of multiple G_1 compartments. Acridine

orange also measuring apoptosis, and for detecting intracellular gradients and the measurement of proton-pump activity (fig.9). The apoptotic cells were greenish yellow (Fig 9 B) in color whereas the necrotic cells were orange-red (Fig.9 A). Significant decrease ($P<0.05$) in apoptotic cells was observed among all epulis whereas maximum apoptosis was observed in giant cell epulis (Fig.9) on interaction which was confirmed by DNA IR absorption bands (Fig.8).

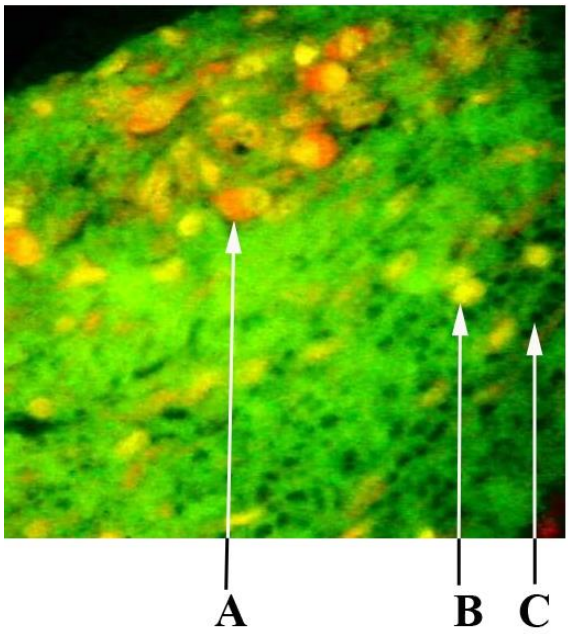

Fig.9 Giant-cell epulis, acridine orange(x800 magnification) A - necrotic cells B - apoptotic cells C - erythrocytes

DISCUSSION

In this study, the expression of MGMT and p53 was examined to elucidate the relationship between their immunostaining. We found that in MGMT cells P53 expression was significantly lower. This is in line with the results of our previous study [28]. Abnormal promoter methylation is not the only determining factor in the regulation of MGMT expression [29]. Other regulators of MGMT expression must be explored to determine this, and the tumor suppressor p53 may be an interesting candidate.

The determination of mutational spectra has defined two areas of contemporary interest in the role of MGMT in mutation avoidance. The first of these is the issue of sequence and DNA strand-specificity of induced mutation. The second concerns the role of MGMT in protection against spontaneous mutation [30].

The recognition of DNA damage (double-strand) start mechanisms leading to apoptosis by activating p53 gene. P53 gene haz its name from the molecular weight of the protein, encoded by it - p53 kDa inactive in normal cells.Now function of the gene P53 can be classified into: helps cell differentiation, cell cycle regulation, cellular responses to DNA damage, DNA repair, start replication and apoptosis. [31].

Abnormal proliferation and various cells damaging is response of activation p53 gene.Cycle cell phase (G1 or G2) stops p53 protein through its target genes. The target gene starts DNA replication before mitosis and stimulates the DNA repair or induces apoptosis. [32, 33]. In addition, the p53 protein itself is involved in DNA repair, inhibits angiogenesis and acts as transcription factor of many genes. [34, 35].

Researches of gene expression TP53 and MGMT in different cell types of human tumors is very inconsistent. Mice and rats given conflicting results about the relationship between the expression of these two genes. Some researchers experimentally demonstrated the inhibitory activity of p53 into MGMT expression, other researchers on cell lines showed that the MGMT gene expression inhibits p53 wild-type. [36-43]Osanai T.et al [44] found that samples of human breast tumors expressing p53 does not inhibit MGMT. Astrocytes cell (mice lines) expressing p53 wild-type have higher activity MGMT. [45]. Decreased MGMT expression found in different samples of human tissue in which the observed changes.in p53. [46].ChakravartiA. et al demonstrated to matched wild-type p53 expression may be necessary for MGMT expression.MGMT and mutant p53 can reduce expression of the genes [47]. Thus, p53 is involved in the regulation of

active MGMT. Summing the results of different studies should indicate that p53 is involved in the induction of gene MGMT and protein activity.However, high levels of expression of p53 suppresses promoter activity of MGMT gene [48].

Owing to the high morphological heterogeneity found in tumors, it has been proposed to use the changes in CH3 and CH2 absorption ratios to avoid normalization of absorptions [49]. In this way, the data treatment of IR images is faster and does not require spectra manipulation to obtain accurate results. However, owing to the weak absorption changes found for phospholipids submitted to oxidative stress, typically a 5–10% absorption decrease for nas(CH2) and a correlative 3–7% absorption increase in n(CH) [50] is observed – global analysis of tumor mass usually fails to highlight these changes between low-grade tumors, for example, astrocytoma and glioblastoma [51].

CONCLUSION.

Aim of this study was to find if IR can be used in neoplasm process DNA methylation detection. Statistical analysis shows that there are differences between spectra of giant cell epulis and control group. Fitting analysis allows to follow small changes in spectra CH3 groups. Presented results prove that infrared spectroscopy could be useful tool in DNA methylation.Morphologically, apoptosis is characterized by DNA fragmentation and formation of apoptotic. DNA CH3 groups can be detected by infrared spectra leads of breaks in DNA strands.This leads to the activation of apoptosis via p53 and MGMT.

Competing interests

The authors declare they have no competing interests

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Romaniuk Anatoly Mykolayovych - ES

Authors' contributions

AB carried out the molecular genetic studies. JY carried out the immunoassays. ES participated in the design of the study and performed the statistical analysis. All authors read and approved the final manuscript.

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Kuzenko Ye. Identification of epulis patients with a high risk of periodontal diseases: DNA infrared spectra, expression MGMT and p53 by and apoptosis analysis / Ye. Kuzenko, A. Romaniuk // Екпериментальна і клінічна медицина. – 2014. - №3 (64). – С.91-102.