MINISTRY OF EDUCATION AND SCIENCE OF UKRAINE SUMY STATE UNIVERSITY MEDICAL INSTITUTE

Eastern Ukrainian Medical Journal

2, Rymskogo-Korsakova st., Sumy 40007, Ukraine e-mail: EUMJ@med.sumdu.edu.ua

eumj.med.sumdu.edu.ua ISSN: 2663-5909 (print)

DOI: https://doi.org/10.21272/eumj.2021;9(1):1-17

Abstract

A. O. Ojetunde,

Ladoke Akintola University of Technology, Ogbomoso, Nigeria

ANTIDIABETIC EFFECTS OF MEDICINAL PLANTS

Diabetes is a chronic disorder that is characterized by an increase in blood glucose (hyperglycemia) with alteration of protein, carbohydrates, and fat metabolism. Consequently, it can lead to renal failure, atherosclerosis, nerve damage, blindness, and coronary heart disease. It is also known as the 5th leading cause of death. Although, there are numerous types of glucose-lowering drugs that exhibit antidiabetic effects but results of treatment in patients are still not so perfect. Therefore, many treatments that include the use of medicinal plants are suggested and encouraged. Medical plants are believed to contain chemical substances with potential curative effects and can often have anti-diabetic effects. This study introduced about 23 effective medicinal plants reported by various experimental researchers with the curative potential to treat diabetes. Although, most of the research used animal models, there is a clear indication that medicinal plants with anti-diabetic potentials are being investigated by several researchers. However, there is a need for further research to be conducted with isolated bioactive ingredients present in these plants in order to have potential ingredients that could be used as a pharmacological agent in the treatment of diabetes mellitus with fewer adverse effects. Again, the mechanisms of action of these medicinal plants in ameliorating diabetes need to be investigated.

Keywords: diabetes mellitus, antidiabetic plants, medicinal plants, hyperglycemia, hypoglycemia, insulin, antioxidants.

Corresponding author: aoojetunde@gmail.com

Introduction

Diabetes is a chronic disorder characterized by an increase in blood glucose (hyperglycemia) with alteration of protein, carbohydrates and fat metabolism [1, 2]. It is caused by either insulin deficiency or malfunction [2]. Diabetes mellitus leads to high blood glucose level, which causes acute complications such as hyperglycemia and hypoglycemia accompanied with long-term complications in many organs of the body, which may result to increased tendency of renal failure, atherosclerosis, nerve damage, blindness and

coronary heart disease resulting in increasing disability [3].

However, it can be classified as either Type I diabetes, Type II diabetes or gestational diabetes mellitus on the basis of clinical representation of the disorder [4]. According to statistics, in 2012, diabetes mellitus affects about 200 million people in the world [5], and it is estimated to rise to over 366 million in the year 2030 [6]. It is also known as the 5th leading cause of death [7]. Oxidative stress (OS) plays a major role in the pathogenesis of micro-vascular and macro-vascular diabetic

1

complications. Hyperglycemia, insulin resistance (IR), and dyslipidemia are present in diabetic patients due to increased OS [8]. Recently, there are different treatments available to control diabetes includes insulin therapy, pharmacotherapy. Additionally, there are numerous types of glucose-lowering drugs that exhibit antidiabetic effects via diverse mechanisms of action [9]. But, inspect of the progress significantly made in the treatment of diabetes, the results of treatment in patients is still not so perfect. These treatments are known to have disadvantages which include side effects, drug resistance (reduction efficiency), and even toxicity [9]. For instance, glucose-lowering drugs are not able to control of excess lipids the presence in blood (hyperlipidemia) [10].

Recently, many treatments that include the use of medicinal plants are suggested and encouraged [11, 12, 13]. Medical plants are believed to contain flavonoids, chemical substances such as carotenoids, terpenoids, glycosides, alkaloids with potential curative effects and can often have antidiabetic effects [14, 15]. The anti-hyperglycemic effects that occur from treatment with plants are most times due to the ability of the plants to improve the action of pancreatic tissue, which is carried out by either increased insulin secretions or reduced intestinal absorption of glucose [9]. The increasing number of people with diabetes has raised a concern for the medical community and the public at large. The aim of this study is to introduce a number of effective medicinal plants with curative potential to treat diabetes and to present other mechanisms of plant compounds used to reduce blood glucose levels. This study will help the medical community and the world at large to find and use a suitable pharmacological agent to treat diabetes with less adverse effects.

Acacia nilotica

nilotica belong to the Mimosaceae. In a study that was carried out to evaluate the efficiency of Acacia nilotica leaf to ameliorate diabetic complications, administration of 50 mg/kg and 200 mg/kg of Acacia nilotica leaf extract to alloxanized mice for 20 days significantly lowered systemic glucose load and insulin resistance without showing any significant effects insulin sensitivity. on Additionally, lowered level of HbA1c improved glucose utilization supported the antihyperglycemic properties of Acacia nilotica leaf. Renal (creatinine, blood urea nitrogen (BUN)) and

hepatic (AST, ALT) and injury markers were was and there normalization dyslipidemia. Furthermore, peroxidase and catalase (CAT) activities in liver, skeletal muscle and kidney increased. It was reported that the plant contains phenolic, flavonoid, catechol, tocopherol and β-sitosterol [16]. In another experiment to determine the effect of Acacia nilotica leaves extract in reducing the high level of glucose and lipid in the blood in alloxanized diabetic rats. Oral administration of Acacia nilotica aqueous methanol leaves extract for 1-3 weeks significantly decreased fasting blood glucose (FBG), low density lipoprotein (LDL), phospholipids, triglyceride (TG), total cholesterol (TC), and very low density lipoprotein (VLDL). In addition, the level of high density lipoprotein (HDL) and the level of serum insulin increased. The duration of these effects was noted after 2 weeks of treatment [8]. Also, 200 mg/kg of Acacia nilotica fruit extract orally administered for 5 weeks to alloxan-induced diabetic rats significantly decreased serum level of LDL and TG, although, there was no significant change in the serum glucose concentrations of diabetic rats [17]. Another study used the Acacia nilotica pods extract to determine its therapeutic effect on streptozotocin (STZ) induced diabetic nephropathy in rat. In this experiment, 150 and 300 mg/kg administration of the extract for 60 days decreased the blood glucose level, and restored and serum The creatinine, urea. normal histopathological architecture of kidney was restored, glomerular size and damaged area were ameliorated, and the adverse effect of diabetes on lipid peroxidation (LPO), superoxide dismutase (SOD) and glutathione (GSH) activity was attenuated [18]. Oral administration of Acacia nilotica leaves extract (300 mg/kg b.w), in comparison to glyburide (900 microgm/kg b.w) for 3 weeks significantly decreased fasting blood glucose FBG and increased insulin level in STZinduced diabetic rats [19]. Also, administration of Acacia nilotica leaves extract resulted into hypoglycemic and anti-platelet aggregation in STZ induced diabetic rats [20]. In summary, aqueous methanol extract from Acacia nilotica (AN) fruits, bark, pods and seeds have been reported to be used traditionally for the treatment of diabetes [19, 33].

Adonsonia digitata

Adonsonia digitata of the family Malvaceae have been reported to be traditionally used for the treatment of diabetes mellitus in Nigeria. The oral administration of the seed prepared by infusion was

2

reported to be used. Also the infusion of the powered fruit of Adonsonia digitata and cow milk was also reported in the study [21]. Traditionally, various parts of the Adonsonia digitata tree have been used to cure many clinical illnesses such as dysentery and diarrhea. Phytochemical screening has indicated that the leaf extract of Adonsonia digitata contains saponins, flavonoids, mucilage, alkaloids and steroids [22]. In an experimental determine and evaluate research to hypolipidaemic and hyperglycaemic effects of methanolic extract of Adonsonia digitata leaves, oral administration of 200 mg/kg and 400 mg/kg of the extract for 6 weeks to STZ induced diabetic rats significantly decreased the blood glucose, TG, cholesterol, glycosylated hemoglobin, LDL, tumor necrosis factor-alpha (TNF-α), interleukin 6 (IL-6), and malondialdehyde (MDA) levels by 46.7%, 43%, 48.91%, 46.15%, 60%, 45.45%, 66% and 30.4%, respectively, in comparison to the diabetic control group. Also, the decline of red blood cell (RBC) count, packed cell volume (PCV), hemoglobin level. HDL and erythropoietin concentration was mitigated. While the level of antioxidant enzymes, SOD and CAT were maintained with reduction in GSH and reduction of elevated white blood corpuscles (WBC) count [22].

Allium cepa

Allium cepa (onions) belonging to the family Amaryllidaceae have been reported to have antidiabetic effects in different experimental researches. A study carried out to evaluate the effects of onion (Allium cepa) dried by heat treatment on FBG level and plasma lipid profile in STZ induced diabetic rats showed a significant lower level of FBG, TG, LDL and TC after 5% onion powered dried at -70°c in a lyophilizator was administered to the diabetic rats. In this experiment HDL level increased [23]. The fasting serum HDL was increased in an experiment carried out to determine the antioxidant activity hypoglycemic effects of Allium cepa in STZ induced diabetic rats. In this experiment, lipoperoxide concentration and lipid hydroperoxide were not increased after administration of Allium cepa [24]. However, in a clinical study with type 1 and type 2 diabetic patients, oral ingestion of crude Allium cepa (100g) by the diabetic patients caused a significant decrease in FBG level in type 1 diabetic patient by about 89 mg/dl in comparison to insulin (145 mg/dl), while in the type 2 diabetic patients, significant decrease in FBG level by 40 mg/dl was observed in relation to glibenclamide (81 mg/dl). In

this same study, after 4 hours, induced hyperglycemia was significantly reduced by about 120 mg/dl in patients with type 1 diabetes, in comparison to water (77 mg/dl) and insulin (153 mg/dl) with same dose of crude Allium cepa, while in type 2 diabetic patient, a significant reduction of about 159 mg/dl compared to water (55 mg/dl) and glibenclamide (114 mg/dl) was observed [25]. Onions is believed to contain flavonoids like quercetin and quercetrin. It also contains sulphur compounds like allyl propyl disulphide and cysteine [25]. These compounds are believed to have antibiotic, anticancer, hypocholesterolaemic, antithrombotic, antioxidant, antibacterial, antidiabetic and fibrinolytic effects [26, 27, 28, 29, 34, 35, 36, 37]. A bioactive flavonoid (Kaempferol-3-O-β-D-6{P- Coumaroyl} Glucopyranoside) that was isolated from Allium cepa was used to determine its antidiabetic effect on alloxan-induced diabetic male rate. The isolated compound administered at 25 mg/kg to the diabetic rats significantly decreased blood glucose in diabetic rats in a manner comparable to the effects obtained with 2 mg/kg of glibenclamide [30]. Also, supplementation of onion powder (7% w/w) administered to STZ-induced diabetic rat for 5 weeks caused reduced level of blood glucose, triglyceride, total serum lipid, renal oxidative stress, and atherogenic index in comparison to the control groups. In this experiment, high density lipoprotein cholesterol/total cholesterol ratio increased [31]. of onion Aqueous extract exhibited antithrombotic effect in STZ-induced diabetic rats

Anacardium occidentalis

Anacardium occidentalis (cashew) belongs to the family Anacardiaceae. The root, bark, stem and leaves of Anacardium occidentalis have been experimentally researched for their antidiabetic effects. For instance, in an experimental study, 2 mg/100g-body weight of crude ethanolic extract of cashew root orally administered to adult guinea pigs and albino rats 3 times daily for 7 days significantly decreased plasma glucose level, cholesterol, liver glycogen and total lipid in guinea pig and albino rats after increased postprandial glucose level, although, plasma protein was not affected [38]. In essence, it could be concluded that ethanolic extract of cashew root can be used as a treatment and for the management of diabetes mellitus. Furthermore, isolated and characterized compounds (stigmast-4-en-3-ol and stigmast-4-en-3-one) gotten from the hexane extract of the bark of

3

Anacardium occidentalis were used in an experiment to determine their hypoglycemic effects. In this experiment, intravenous administration of these compounds at 1.3 mg/kg b.w significantly reduced blood glucose level in normal, healthy dogs. It was hypothesized that these compounds presence in cashew bark could be responsible for its hypoglycemic effects [39]. Also, of 100 mg/kgAnacardium occidentalis administered to alloxan-induced diabetic rats showed a significant increase in SOD activity [40]. In another experiment, when graded doses (100-800 mg/kg p.o.) of aqueous and methanolic stembark extracts of A. occidentale was administered to STZ induced diabetic rat, there was dosedependent, significant decrease in the blood glucose level of fasted normal and fasted diabetic rats. In the same study, a single dose of 800 mg/kg p.o., of Anacardium occidentalis stem-bark aqueous and methanolic extracts significantly decreased mean basal blood glucose level of fasted normal and fasted diabetic rats, although, it was reported that these extract is less potent in comparison to insulin. The authors concluded that the presence of terpenoid and/or coumarin in the extract could have caused the hypoglycemic effects but the mechanism is not yet fully understood [41]. In another study, administration of methanolic leaves extract of Anacardium occidentalis to alloxan-induced diabetic rat caused 79.2% change compared to Tolbutamide (63.1%) over 4 hours for moderately diabetic rat. In the experiment, when diabetes became severe, the extract reduced blood glucose level by 20.8% compared to Tolbutamide (47.63%) over 4 hours, although, the values were not considered to be significant. So, Anacardium occidentalis is believed to have similar ability in lowering blood glucose concentration compared to Tolbutamide (a reference drug) [42]. Injection of 100 mg/kg of Anacardium occidentalis plant extract to neonatal STZ diabetic rats for 30 days caused a significant decrease in FBG level. The effects gotten are similar to the treatment with Pioglitazone (a standard drug). It was reported that, for future purpose, specific compound(s) responsible for the antidiabetic effects of Anacardium occidentalis is needed to be investigated [49].

Azadirachta indica

Neem (A. indica) belongs to the family Meliaceae. Traditionally, maceration of Azadirachta indica leaves and Vernonia amygdalina (bitter leaf) has been reported to be orally used to treat diabetes in Nigeria [21].

Azadirachta indica is known to have hypoglycemic, hepatoprotective hypolipidemic, and immunostimulant properties [43, 44]. Some chemical compounds such as nimolinone, nimbocinone, kulactone, isonimocinolide. nimocinolides, nimbin, azadirachtin, salanin, flavonoids, meldenindiol, myricetin, isomargosinolide, margosinolide, desacetyldihydronimbic acid and vilasinin have been isolated from A. indica leaves [45, 46, 47]. In an experimentally study, ethanolic extract of Vernonia amygdalina (VA) and Azadirachta indicia (AI) co-administered at 200 mg/kg, 50:50 to STZ-induced diabetic rats for 28 days reduced blood glucose, T3 and T4. Decreased glutathione peroxidase (GPx) and CAT activities were ameliorated and SOD activities increased. It was reported that the antidiabetic synergistic action of VA/AI could be due to insulin mimetic action, oxidative stress attenuation and βregeneration [48]. In another study, intraperitoneal injection of chloroform plant extract of Azadirachta indicia to STZ induced diabetic mice for 21 days regenerated insulin-producing cells with increase in plasma insulin and c-peptide levels. In the experiment, intestinal glucosidase activity reduced. glucose-6-phosphate dehydrogenase activity and hepatic, skeletal muscle glycogen content increased, and oral glucose was well tolerated [50]. A. indicia is reported to have bioactive compounds such as rutin, quercetin and nimbidin, which is said to be responsible for its hypoglycemic effects [51]. The hypoglycemic effect of combination of Azadirachta indicia and Gynura procumbens was carried out by Sunarwidhi et al. In the experiment, the macerated extracts administered to alloxan-induced diabetic rats for 15 days significantly improved the morphology of βcells and the islets of Langerhans. Also, insulin and expression increased elevated-glucose concentration decreased [51]. Furthermore, in a randomized, double-blind, placebo-controlled clinical study, subjects with type 2 diabetes already placed on standard metformin therapy received different doses of neem for 12 weeks. In this experiment, neem at 125, 250, and 500mg doses significantly decreased postprandial blood sugar level, FBG, HbA1C and IR compared to the placebo group. Endothelial function improved OS and systemic inflammation decreased but there was no effects on lipid profile or platelet aggregation. It was suggested by the authors that neem may cure systemic inflammation hyperglycemia, endothelial dysfunction in patients with type 2

diabetes in comparison to the effects of metformin [52]. In another study, butanol fraction of Azadirachta indica ethanol stem bark extract showed DDPH scavenging activity, FRAP activity, ameliorated oxidative injury in hepatic tissue by reducing malondialdehyde (MDA) concentration significantly, improved the activities of SOD and CAT, improved glucose uptake in psoas muscle with or without insulin, and inhibited activities of α-glucosidase α -amylase. Sistosterol, and campestrol, stigmasterol, squalene and nimbiol are reported to be present in Azadirachta indica. It was suggested that butanol and ethyl acetate fractions of A. indica may possibly have bioactive compounds with potentials to cure diabetes [53]. Impaired nerve functions and delayed nerve recovery occurring due to hyperglycemia-induced OS was ameliorated in STZ-induced diabetic rats by the action of A. indica flower extract at a dose of 250. 500 or 750 mg/kg. In this experiment, functional recovery (motor and sensory functions) improved significantly, MDA levels significantly decreased, while SOD activity and axon density significantly increased. It was suggested by the authors that A. indica flower extract may have antioxidative effect [54]. In another study, combination of (1:1) aqueous extract of dried powder of Azadirachta indica (leaves) and Abroma augusta (root) orally administered to alloxan-induced diabetic rats once a day for 8 weeks significantly decrease blood sugar, serum lipids, formation of lipid peroxides and LPO antioxidants (SOD, increased glutathione transferase and glutathione peroxidase). Decrease in body weight was also prevented by the extract [55]. Also, chloroform leaf extract of Azadirachta indica increased GSH, SOD, CAT and oxidized glutathione (GSSG), hepatic glycogen content, insulin plasma and glucose-6-phosphatase in STZ-induced diabetic rats after chronic oral administration of the extract for 28 days. Meanwhile, IR, lipid peroxidation and glucokinase (GK) decreased. It was reported that A. indica can be considered to be a potential antidiabetic-safe agent [56]. Absence of marked hyperglycemia, absence of diabetic nephropathy, absence of glomerulosclerosis and absence nodular vacuolation of proximal tubule cells was observed in an experiment carried out to determine the ameliorative effects of ethanolic leaf extract of Azadirachta indica (500 mg/kg b.w) for 50 days on renal histological alterations in STZ-induced diabetic rats. It was stated that leaf extract of

Azadirachta indica ameliorates hyperglycemia and diabetic nephropathy in rats [57].

Balanites aegyptiaca

Balanites aegyptiaca commonly known as desert date tree belongs to the family Balanitiaceae. The kernel fruit was reported to contain polyphenols [58] and saponins [59]. Other bioactive compounds like flavonoids, alkaloids, tannins and vitamins have been reported in the fruit as well as the branches, leaves and roots of Balanites aegyptiaca [60, 61, 62]. In a study, 50 mg/kg b.w of crude extract, butanol or dichloromethane fraction of Balanites aegyptiaca administered to diabetic rats produced a decrease in plasma glucose, lactic acid, HbA1c, lipid profile, MDA, GSH levels, CAT and SOD activities with an increase in insulin and insulin receptor substrate 1 in rat pancreas. In essence, it was suggested by the authors that the hypoglycemic effect of Balanites aegyptiaca is due to the inhibition of the SAPK-JNK pathway [63]. In another study, B. aegyptia significantly decreased mean plasma glucose and MDA levels and significantly increased mean plasma insulin, total antioxidant capacity (TAC) levels, and liverpyruvate kinase (L-PK) in STZ-induced diabetic rats after oral administration of Balanites aegyptiaca fruits aqueous extract (1.5 g/kg b.w) daily for 45 days. Size of the islets of Langerhans and weight of the pancreas increased and histoarchitecture also improved [64]. In a randomized double-blinded pilot clinical study to determine the antidiabetic efficacy of 70% ethanol extract of the pericarps of B. aegyptiaca on type 2 diabetic patients, B. aegyptiaca incorporation in hard gelatine capsules and administration at 400 mg/day for 8 weeks decreased postprandial plasma glucose, FBG, TG, LDL, TC, AST, ALT significantly. In the experiment, HDL increased. Administration of *B. aegyptiaca* capsules to type 2 diabetic patients caused significant improvements in glycaemic markers and lipid profile, without adverse effects or hypoglycemia [65]. Ethyl acetate extract from Balanites aegyptiaca administered at 10, 20 or 50 mg/kg b.w to experimental diabetic rats for 8 weeks lowered blood glucose level, HbA1c, MDA and vascular endothelial growth factor (VEGF) in diabetic retina. Tumor necrosis factor alpha (TNF-α) and interleukin (IL-1β) significantly decreased in diabetic rats treated with the extract and β-sistosterol was present in the extract [66].

Brassica oleracea

Brassica oleracea (broccoli) belongs to the family Brassicaceae. It is said to contain

5

, 2021

components like minerals, vitamins, dietary fiber, hydroxycinnamic acids, flavonol glycosides and glucosinolates [67]. In an experimental study, when STZ-induced diabetic rats were administered polyphenols (5 mL/week) gotten from aqueous broccoli extract for 8 weeks, DNA damage reduced TAC significantly, GSH and values were significantly conserved, and pancreatic histopathological changes were attenuated. The author concluded that B. oleracea reduced the STZ mediated hyperglycemia and the STZ-induced oxidative injury to pancreas tissue [67]. In a recent study, a single oral administration of aqueous extract of B. oleracea at a dose of 60 mg/kg significantly decreased blood glucose at the 6th hour in STZ-induced diabetic rats. In the study, repeated administration of the same dose for 7 days significantly decreased the blood glucose to the normal level. The author revealed that B. oleracea aqueous extract is rich in numerous phytochemical compounds and can exert antioxidant activity [68].

In another study, 500 mg/kg b.w of B. oleracea methanol extract administered to alloxan-induced diabetic rats significantly lowered FBG, TC, and LDL, whereas the HDL increased in comparison to the diabetic control group. The changes were said to be similar in comparison to glibenclamide (a reference drug) [69]. However, the antioxidant activity B. oleracea edible sprouts [70], it phytochemical components [71], and amino acid compositions [72] have been reported. Furthermore, 800 mg/kg b.w of B. oleracea aqueous extract administered to STZ-induced diabetic rats for 28 days significantly decreased FBG by about 64% within 7 days of treatment. Additionally, HbA1c and lipid profile normalized. The authors also declared that BUN, Serum glutamic oxaloacetic transaminase (SGOT) and Serum glutamic pyruvic (SGPT) significantly transaminase decreased, meanwhile, activities of CAT and SOD significantly increased. However, chlorogenic acid, sinapic acid and rutin were present in the extract [73].

Table 1 – Other medicinal plants with investigated antidiabetic effects

No.	Botanical name	Family	Common name	Part(s) used	Significant antidiabetic activities
1	Carica papaya	Caricaceae	Pawpaw	Leaves, fruits	Preserved integrity of pancreatic islets, improved basal insulin secretion and protected cultured call from adverse effects of STZ [74]. Exhibited hypoglycemic and antioxidant effects and improved lipid profile [75]. Improved platelet function and increased total antioxidant capacity (TAC) and SOD in type 2 diabetic patients [76]. Decreased blood glucose and serum lipid levels [77, 78, 79, 38].
2	Eugenia caryophyllus	Myrtaceae	Clove	Bud	Inhibited α -amylase and α -glucosidase activities and exhibited antioxidants activities [80].
3	Ficus carica	Moraceae	Fig	Fruit	Improved cholesterolaemic status [81]. Regulated blood glucose and lipids parameters [82]. Lowered blood glucose, TC, and TG to normal [83].
4	Ficus deltoidea	Moraceae	Fig	Leaf	Stimulated insulin secretion and blocked the production of hepatic glucose [84]. Suppressed hepatic glucose output, improved insulin sensitivity and enhanced glucose uptake in type 2 diabetes mellitus [85]. Decreased total and LDL-c concentration [86]. Decreased blood glucose to near normal [100, 101]. Promoted regeneration of islet, increased antioxidant enzymes of pancreas and increased insulin secretion [101].
5	Ficus racemosa	Moraceae	Cluster fig, redwood fig	Bark, stem, leaves, root	Decreased blood glucose [87, 88, 92, 93, 94 95], serum lipid, and lipoprotein [89], serum cholesterol, serum triglycerides and serum urea [90]. Exhibited increased in plasma insulin level [91] and inhibited the activity of hexokinase and glucose 6-phosphatase [89].

No.	Botanical name	Family	Common name	Part(s) used	Significant antidiabetic activities
6	Ficus thonningii	Moraceae	Wild fig	Stem bark	Exhibited hypoglycemic effects [96, 97, 98], and hypolipidaemic effects [99]. Increased glucose uptake in primary hepatocytes [99].
7	Gossypium herbaceum	Malvaceae	Cotton	Seed	Reduced serum level of glucose, TG, cholesterol, urea and creatinine [102].
8	Guiera senegalensis	Combretaceae	Sabara	Leaves and root	Increased body weight and HDL-c, and decreased glycaemia, insulin, LDL-c, TG, TC creatinine and urea [103].
9	Khaya senegalensis	Meliaceae	Mahogany	Root, stem, bark	Reduced the level of blood glucose, stimulated synthesis of hepatic glycogen, improved tolerance of oral glucose and function of β -cell, decreased insulin resistance, ameliorated alterations of serum lipids and prevented renal and hepatic damages [104]. Inhibited α -glucosidase and α -amylase activities [105, 106].
10	Lawsonia inermis	Lythraceae	Egyptian priest, henna	Leaves	Decreased glucose, cholesterol, and TG concentration to normal [107]. Improved plasma albumin, lipid profile, serum creatinine and total plasma protein [108].
11	Mangifera indica	Anacardiaceae	Mango	Leaves, kernel flour.	Exhibited dose-dependent inhibition against α-glucosidase activities [109, 110] and α-amylase [110]. Decreased blood glucose level beyond glibenclamide effects with increase in the sensitivity of insulin and plasma insulin levels [111]. Prevented the decline in body weight and decrease in β-cell mass [112]. Improved FBG, HbA1c, hepatic glycogen, plasma electrolytes, lipid profile, pancreatic and hepatic MDA, and the markers of liver function [113].
12	Moringa oleifera	Moringaceae	Drumstick	Leaf, seed, fruit	Significantly decreased blood glucose in diabetic rats and mice [114-130]. Reduced triglycerides levels [115]. Increased CAT [116, 119, 130], increased SOD [118. 119. 130] and decreased MDA [116, 118, 119]. Increased HDL [120, 121, 122], decreased cholesterol, LDL, VLDL and triglycerides [120, 121, 122, 125]. Decreased HbA1c level [126, 129].
13	Parkia biglobosa	Fabaceae	Locust bean	Seed	Decreased FBG [132, 133] cholesterol, serum triglyceride, LDL-c, VLDL cholesterol and LPO with increase in HDL-c and restoration of biomarkers of OS [131]. Improvement of glucose tolerance and pancreatic β-cell function with stimulation of insulin secretion, decrease in insulin resistance, restoration of liver glycogen amelioration of serum dyslipidaemia and prevention of renal and hepatic damages in comparison to the untreated diabetic rats [132].
14	Psidium guajava	Myrtaceae	Guava	Leaves	Decreased TC [134, 135 137, 140], LDL [134, 135, 137, 140], glucose level [134, 136-141], and TG [135, 137, 140]. Increased plasma insulin level [138], HDL [135, 137, 140], SOD and CAT activity [141].
15	Solanum incanum	Solanaceae	Bitter apple	Fruit	Reduced blood glucose concentration [142].

No.	Botanical name	Family	Common name	Part(s) used	Significant antidiabetic activities
16	Vernonia amygdalina	Asteraceae	Bitter leaf	Leaves	Exhibited antihyperglycemic effects, decreased LDL-c, VLDL cholesterol and increased HDL-c in diabetic rats [143]. Improved glucose tolerance, decreased FBG, TG and TC, protected β-cells and increased insulin in diabetic rats [144]. Decreased TG and MDA levels and normalized cholesterol concentration [145].
17	Ziziphus mucronata	Rhamnaceae	Buffalo thorn	Root,	Lowered blood glucose, improved glucose tolerance, and increased serum insulin and liver glycogen [146].

Abbreviations: streptozotocin (STZ); total cholesterol (TC); triglycerides (TG); low density lipoprotein cholesterol (LDL-c); high density lipoprotein (HDL); low density lipoprotein (LDL), very low density lipoprotein (VLDL); superoxide dismutase (SOD); catalase (CAT); malondialdehyde (MDA); glycosylated hemoglobin (HbA1c); high density lipoprotein cholesterol (HDL-c); oxidative stress (OS); lipid peroxidation (LPO); fasting blood glucose (FBG)

Conclusions

In summary, this present study has listed some medicinal plants with reported and potential antidiabetic effects. Although, most of the research used animal models, there is a clear indication that medicinal plants with anti-diabetic potentials are being investigated by several researchers. Unfortunately, most of the investigations are preliminary in nature. However, there is a need for further research to be conducted with isolated

bioactive ingredients present in these plants in order to have potential compounds that could be used as a pharmacological agent in the treatment of diabetes mellitus. Again, the mechanisms of action of these medicinal plants in ameliorating diabetes need to be investigated. Moreover, pharmaceutical industries need to support more research activities in this area in order to produce and commercially utilize antidiabetic product from medicinal plants with less disadvantages/adverse effects.

References (список літератури)

- Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TP. Indian herbs and herbal drugs used for the treatment of diabetes. *J Clin Biochem Nutr*. 2007;40(3):163-173. doi:10.3164/jcbn.40.163.
- Osadebe PO, Odoh EU, Uzor PF. The search for new hypoglycemic agents from plant. *Afr J Pharm Pharmacol*. 2014;8(11):292-303. doi: 10.5897/AJPP2014.3933.
- 3. American Diabetes Association. 2. Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes-2019. Diabetes Care. 2019;42(1):13-28. doi:10.2337/dc19-S002.
- Velho G, Froguel P. Maturity-onset diabetes of the young (MODY), MODY genes and non-insulin-dependent diabetes mellitus. *Diabetes Metab*. 1997;23 Suppl 2:34-37.
- 5. Wais M, Nazish I, Samad A, Beg S, Abusufyan S, Ajaj SA, et al. Herbal drugs for diabetic treatment: an updated review

- of patents. *Recent Pat Antiinfect Drug Discov*. 2012;7(1):53-59. doi:10.2174/157489112799829701.
- 6. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care*. 2004;27(5):1047-1053. doi:10.2337/diacare.27.5.1047.
- 7. Kazi S. Use of traditional plants in diabetes mellitus. *Int J Pharm.* 2014;4(4):283-289.
- 8. Asad M, Munir TA, Farid S, Aslam M, Shah SS. Duration effect of Acacia nilotica leaves extract and glibenclamide as hypolipidaemic and hypoglycaemic activity in alloxan induced diabetic rats. *J Pak Med Assoc*. 2015;65(12):1266-1270.
- Kooti W, Farokhipour M, Asadzadeh Z, Ashtary-Larky D, Asadi-Samani M. The role of medicinal plants in the treatment of diabetes: a systematic review. *Electron Physician*. 2016;8(1):1832-1842. doi:10.19082/1832.

10. Dey L, Attele AS, Yuan CS. Alternative therapies for type 2 diabetes. *Altern Med Rev.* 2002;7(1):45-58. PMID: 11896745.

- Kooti W, Moradi M, Akbari SA, Sharafi-Ahvazi N, AsadiSamani M, Ashtary-Larky
 D. Therapeutic and pharmacological potential of Foeniculum vulgare Mill: A review. *J Herb Med Pharmacol*. 2015;4:1-9.
- 12. Okwu DE. Evaluation of the chemical composition of indigenous spices and flavouring agents. *Global J Pure Appl Sci.* 2001;7(3):455-459. doi:10.4314/gjpas.v7i3.16293.
- 13. Newman DJ. Natural products as leads to potential drugs: an old process or the new hope for drug discovery? *J Med Chem.* 2008;51(9):2589-2599. doi:10.1021/jm0704090.
- Afrisham R, Aberomand M, Ghaffari MA, Siahpoosh A, Jamalan M. Inhibitory effect of Heracleum persicum and Ziziphus jujuba on activity of Alpha-Amylase. *Journal of Botany*. 2015;2015:1-8. doi:10.1155/2015/824683.
- Kuhn M, Winston D. Herbal therapy and supplements: a scientific and traditional approach. New York: Lippincott and Wilkins, 2000;347-350.
- 16. Saha MR, Dey P, Sarkar I, Sarker DD, Haldar B, Chaudhuri TK, et al. Acacia nilotica leaf improves insulin resistance and hyperglycemia associated acute hepatic injury and nephrotoxicity by improving systemic antioxidant status in diabetic mice. *J Ethnopharmacol*. 2018;210:275-286. doi:10.1016/j.jep.2017.08.036.
- 17. Abuelgassim AO. Effect of Acacia nilotica fruit extract on serum glucose and lipid concentrations in alloxan-induced diabetic rats. *Pak J Biol Sci.* 2013;16(21):1398-1402. doi:10.3923/pjbs.2013.1398.1402.
- 18. Omara EA, Nada SA, Farrag AR, Sharaf WM, El-Toumy SA. Therapeutic effect of Acacia nilotica pods extract on induced streptozotocin diabetic rat. nephropathy in Phytomedicine. 2012;19(12):1059-1067. doi:10.1016/j.phymed.2012.07.006.
- Asad M, Munir TA, Afzal N. Acacia nilotica leave extract and glyburide: comparison of fasting blood glucose, serum insulin, beta-thromboglubulin levels

- and platelet aggregation in streptozotocin induced diabetic rats. *J Pak Med Assoc*. 2011;61(3):247-251.
- 20. Asad M, Munir TA, Afzal N. Acacia nilotica leave extract and glyburide: comparison of fasting blood glucose, serum insulin, beta-thromboglubulin levels and platelet aggregation in streptozotocin induced diabetic rats. *J Pak Med Assoc*. 2011;61(3):247-251.
- Abubakar US, Abdullahi S, Ayuba V, Kaigama S, Halidu US, Ayuba MK. Medicinal plants used for the management of diabetes mellitus in Zaria, Kaduna state, Nigeria. *Journal of Pharmacy & Pharmacognosy Research*. 2017;5(3):156-164.
- 22. Ebaid H, Bashandy SAE, Alhazza IM, Hassan I, Al-Tamimi J. Efficacy of a methanolic extract of *Adansonia digitata* leaf in alleviating hyperglycemia, hyperlipidemia, and oxidative stress of diabetic rats. *Biomed Res Int*. 2019;2019:2835152. doi:10.1155/2019/2835152.
- 23. Ülger TG, Çakiroglu FP. The effects of onion (*Allium cepa* L.) dried by different heat treatments on plasma lipid profile and fasting blood glucose level in diabetic rats. *Avicenna J Phytomed*. 2020;10(4):325-333.
- 24. Campos KE, Diniz YS, Cataneo AC, Faine LA, Alves MJ, Novelli EL. Hypoglycaemic and antioxidant effects of onion, Allium cepa: dietary onion addition, antioxidant activity and hypoglycaemic effects on diabetic rats. *Int J Food Sci Nutr.* 2003;54(3):241-246. doi:10.1080/09637480120092062.
- 25. Taj Eldin IM, Ahmed EM, Elwahab H M A. Preliminary Study of the clinical hypoglycemic effects of Allium cepa (Red Onion) in Type 1 and Type 2 diabetic patients. *Environ Health Insights*. 2010;4:71-77. doi:10.4137/EHI.S5540.
- 26. Dorant E, van den Brandt PA, Goldbohm RA. Allium vegetable consumption, garlic supplement intake, and female breast carcinoma incidence. *Breast Cancer Res Treat*. 1995;33(2):163-170. doi: 10.1007/BF00682723.

27. Banerjee SK, Maulik SK. Effects of garlic on cardiovascular disorders: a review. *Nutritional Journal*. 2002;1(4):1–14.

- 28. Carson JF. Chemistry and biological properties of onion and garlic. *Food Reviews Internationl.* 1987;3:71-103.
- 29. Reddy BS, Rao CV, Rivenson A, Kelloff G. Chemoprevention of colon carcinogenesis by organosulfur compounds. *Cancer Res.* 1993;53(15):3493-3498.
- 30. Ikechukwu OJ, Ifeanyi OS. The antidiabetic effects of the bioactive flavonoid (Kaempferol-3-O-β-D-6{P-Coumaroyl} Glucopyranoside) isolated from Allium cepa. *Recent Pat Antiinfect Drug Discov*. 2016;11(1):44-52. doi:10.2174/1574891x11666151105130233.
- 31. Bang MA, Kim HA, Cho YJ. Alterations in the blood glucose, serum lipids and renal oxidative stress in diabetic rats by supplementation of onion (Allium cepa. Linn). *Nutr Res Pract*. 2009;3(3):242-246. doi:10.4162/nrp.2009.3.3.242.
- 32. Jung YS, Kim MH, Lee SH, Baik EJ, Park SW, Moon CH. Antithrombotic effect of onion in streptozotocin-induced diabetic rat. *Prostaglandins Leukot Essent Fatty Acids*. 2002;66(4):453-458. doi:10.1054/plef.2002.0373.
- 33. Sundaram R, Mitra SK. Antioxidant activity of ethyle acetate soluble fraction of Acacia Arabica barks in rats. *Indian J Pharmacol.* 2007;39:33-38.
- 34. Torres-Urrutia C, Guzmán L, Schmeda-Hirschmann G, et al. Antiplatelet, anticoagulant, and fibrinolytic activity in vitro of extracts from selected fruits and vegetables. *Blood Coagul Fibrinolysis*. 2011;22(3):197-205. doi:10.1097/MBC.0b013e328343f7da.
- 35. Yamada K, Naemura A, Sawashita N, Noguchi Y, Yamamoto J. An onion variety has natural antithrombotic effect as assessed by thrombosis/thrombolysis models in rodents. *Thromb Res*. 2004;114(3):213-220. doi:10.1016/j.thromres.2004.06.007.
- 36. Yamamoto Y, Aoyama S, Hamaguchi N, Rhi GS. Antioxidative and antihypertensive effects of Welsh onion on rats fed with a high-fat high-sucrose diet. Biosci Biotechnol Biochem.

- 2005;69(7):1311-1317. doi:10.1271/bbb.69.1311.
- Kabrah MA, Faidah HS, Ashshi AM, Turkistani MSA. Antibacterial Effect of Onion. Sch J App Med Sci. 2016;4:4128-4133.
- 38. Egwim E. Hypoglycemic potencies of crude ethanolic extracts of cashew roots and unripe pawpaw fruits in guinea pigs and rats. *J Herb Pharmacother*. 2005;5(1):27-34.
- 39. Alexander-Lindo RL, Morrison EY, Nair MG. Hypoglycaemic effect of stigmast-4-en-3-one and its corresponding alcohol from the bark of Anacardium occidentale (cashew). *Phytother Res.* 2004;18(5):403-407. doi:10.1002/ptr.1459.
- 40. Okpashi VE, Bayim BP, Obi-Abang M. Comparative effects of some medicinal plants: Anacardium occidentale, Eucalyptus globulus, Psidium guajava, and Xylopia aethiopica extracts in alloxaninduced diabetic male wistar albino rats. *Biochem Res Int.* 2014;2014:203051. doi:10.1155/2014/203051.
- 41. Ojewole JA. Laboratory evaluation of the hypoglycemic effect of Anacardium occidentale Linn (Anacardiaceae) stembark extracts in rats. *Methods Find Exp Clin Pharmacol*. 2003;25(3):199-204. doi:10.1358/mf.2003.25.3.769640.
- 42. Fagbohun TR, Odufuwa KT. Hypoglycemic effect of methanolic extract of Anacardium occidentale leaves in alloxan-induced diabetic rats. *Niger J Physiol Sci.* 2010;25(1):87-90.
- 43. Khosla P, Bhanwara S, Singh, J, Seth S, Srivastava RK. A study of hyperglycemia effects of *A. indica* (Neem) in normal and alloxan diabetic rabbits. *Indian Journal of Physiology & Pharmacology*. 2000;44:69–74.
- 44. Bopana KN, Kannan J, Gadgil S, Balaram R Rathod SP. Antidiabetic and antihyperlipidaemic effects of neem seed kernel powder on alloxan diabetic rabbits. *Indian Journal of Pharmacology*. 1997;29(3):162–16.
- 45. Govindachari TR, Sandhya G, Ganeshraj SP. Simple method for the isolation of azadirachtin by preparative high-performance liquid chromatography. *Journal of Chromatography*.

- 1990;513:389–391. doi:10.1016/S0021-9673(01)89462-0.
- 46. Ara I, Siddiqui BS, Faizi S Siddiqui S. Diterpenoids from the stem bark of *Azadirachta indica*. *Phytochemistry*. 1989;28(4):1177–1180. doi:10.1016/0031-9422(89)80204-3.
- Basak SP, Chakroborty DP. Chemical investigation of *Azadirachta indica* leaf (M. azadirachta). *Journal of the Indian Chemical Society*. 1969;45:466–467.
- 48. Atangwho IJ, Ebong PE, Eyong EU, Asmawi MZ, Ahmad M. Synergistic activity of antidiabetic Vernonia amygdalina and Azadirachta indica: biochemical effects and possible mechanism. JEthnopharmacol. 2012;141(3):878-887. doi:10.1016/j.jep.2012.03.041.
- 49. Jaiswal YS, Tatke PA, Gabhe SY, Vaidya AB. Antidiabetic activity of extracts of *Anacardium occidentale* Linn. leaves on *n*-streptozotocin diabetic rats. *J Tradit Complement Med*. 2016;7(4):421-427. doi:10.1016/j.jtcme.2016.11.007.
- 50. Bhat M, Kothiwale SK, Tirmale AR, Bhargava SY, Joshi BN. Antidiabetic properties of Azardiracta indica and Bougainvillea spectabilis: In vivo Studies in murine diabetes model. *Evid Based Complement Alternat Med*. 2011;2011:561625. doi:10.1093/ecam/nep033.
- 51. Sunarwidhi AL, Sudarsono S, Nugroho AE. Hypoglycemic effect of combination of Azadirachta indica A. Juss. and Gynura procumbens (Lour.) Merr. Ethanolic Extracts Standardized by Rutin and Quercetin in Alloxan-induced Hyperglycemic Rats. Adv Pharm Bull. 2014 Dec;4(2):613-618. doi: 10.5681/apb.2014.090.
- 52. Pingali U, Ali MA, Gundagani S, Nutalapati C. Evaluation of the effect of an aqueous extract of *Azadirachta indica* (Neem) leaves and twigs on glycemic control, endothelial dysfunction and systemic inflammation in subjects with Type 2 diabetes mellitus A randomized, double-blind, placebo-controlled clinical study. *Diabetes Metab Syndr Obes*. 2020;13:4401-4412. doi:10.2147/DMSO.S274378.

- 53. Sanni O, Erukainure OL, Chukwuma CI, Koorbanally NA, Ibeji CU, Islam MS. Azadirachta indica inhibits key enzyme linked to type 2 diabetes in vitro, abates oxidative hepatic injury and enhances muscle glucose uptake ex vivo. *Biomed Pharmacother*. 2019;109:734-743. doi:10.1016/j.biopha.2018.10.171.
- 54. Sriraksa N, Kongsui R, Thongrong S, Duangjai A, Hawiset T. Effect of *Azadirachta indica* flower extract on functional recovery of sciatic nerve crush injury in rat models of DM. *Exp Ther Med*. 2019;17(1):541-550.
 - doi: 10.3892/etm.2018.6931.
- 55. Halim EM. Lowering of blood sugar by water extract of Azadirachta indica and Abroma augusta in diabetes rats. *Indian J Exp Biol.* 2003;41(6):636-640.
- Gutierrez RM, Gómez YG, Guzman MD. Attenuation of nonenzymatic glycation, hyperglycemia, and hyperlipidemia in streptozotocin-induced diabetic rats by chloroform leaf extract of Azadirachta indica. *Pharmacogn Mag*. 2011;7(27):254-259. doi:10.4103/0973-1296.84243.
- 57. Oluwole BA, Laura Z, Olufunke OD, Oluwafunmike SA, Luciana D, Ezekiel CM. Ameliorative effects of ethanolic leaf extract of Azadirachta indica on renal histologic alterations in streptozotocininduced diabetic rats. *Am J Chin Med*. 2011;39(5):903-916. doi:10.1142/S0192415X11009299.
- 58. Ahmed AA, Kita A, Nem's A, Miedzianka J, Foligni R, Abdalla AM, et al. Tree-to-tree variability in fruits and kernels of a Balanites aegyptiaca (L.) Del. population grown in Sudan. *Trees*. 2019;34(1) doi:10.1007/s00468-019-01901-x.
- 59. Yadav JP, Panghal M. Balanites aegyptiaca (L.) Del. (Hingot): A review of its traditional uses, phytochemistry and pharmacological properties. *Int. J. Green Pharm.* 2010;4(3):140–146. doi: http://dx.doi.org/10.22377/ijgp.v4.
- 60. Maksoud SA, El Hadidi M.N. The flavonoids of Balanites aegyptiaca (Balanitaceae) from Egypt. *Plant Syst. Evol.* 1988;160:153–158. doi: https://doi.org/10.1007/BF00936042.
- 61. Sagna MB, Diallo A, Sarr PS, Ndiaye O, Goner D, Guisse A. Biochemical

composition and nutritional value of Balanites aegyptiaca (L.) Del fruit pulps from Northen Ferlo in Senegal. *Afr. J. Biotechnol.* 2014;13(2):336–342.

- 62. Farid H, Haslinger E, Kunert O. New steroidal glycosides from Balanites aegyptiaca. *Helv. Chim. Acta.* 2002;85(4):1019–1026. doi:10.1002/1522-2675(200204)85:4%3C1019::AID-HLCA1019%3E3.0.CO;2-S.
- 63. Hassanin KMA, Mahmoud MO, Hassan HM, Abdel-Razik AH, Aziz LN, Rateb ME. Balanites aegyptiaca ameliorates insulin secretion and decreases pancreatic apoptosis in diabetic rats: Role of SAPK/JNK pathway. *Biomed Pharmacother*. 2018;102:1084-1091. doi:10.1016/j.biopha.2018.03.167.
- 64. Abou Khalil NS, Abou-Elhamd AS, Wasfy SI, El Mileegy IM, Hamed MY, Ageely HM. Antidiabetic and antioxidant impacts of desert date (Balanites aegyptiaca) and parsley (Petroselinum sativum) aqueous extracts: Lessons from experimental rats. *J Diabetes Res.* 2016;2016:8408326. doi:10.1155/2016/8408326.
- 65. Rashad H, Metwally FM, Ezzat SM, Salama MM, Hasheesh A, Abdel Motaal A. Randomized double-blinded pilot clinical study of the antidiabetic activity of Balanites aegyptiaca and UPLC-ESI-MS/MS identification of its metabolites. *Pharm Biol.* 2017;55(1):1954-1961. doi:10.1080/13880209.2017.1354388.
- 66. Al-Malki AL, Barbour EK, Abulnaja KO, Moselhy SS. Management of hyperglycaemia by ethyl acetate extract of Balanites aegyptiaca (Desert Date). *Molecules*. 2015;20(8):14425-14434. doi:10.3390/molecules200814425.
- 67. Suresh S, Waly MI, Rahman MS, et al. Broccoli (*Brassica oleracea*) reduces oxidative damage to pancreatic tissue and combats hyperglycaemia in diabetic rats. *Prev Nutr Food Sci.* 2017;22(4):277-284. doi:10.3746/pnf.2017.22.4.277.
- 68. Amssayef A, Eddouks M. Antihyperglycemic effect of the moroccan collard green (Brassica oleracea var. viridis) in streptozotocin-induced diabetic rats published online ahead of print, 2020 Sep 29. Endocr Metab Immune Disord Drug Targets.

- 2020;10.2174/1871530320666200929141140 .doi:10.2174/1871530320666200929141140.
- Assad T, Khan RA, Feroz Z. Evaluation of hypoglycemic and hypolipidemic activity of methanol extract of Brassica oleracea. *Chin J Nat Med.* 2014;12(9):648-653. doi:10.1016/S1875-5364(14)60099-6.
- 70. Lim JH, Park KJ, Jeong JW, Park JJ, Kim BK, Kim JC, et al. Antioxidant activity and antioxidant compounds in edible sprouts. *FASEB J* 27: lb260 2013
- Baenas N, Moreno DA, García-Viguera C. Selecting sprouts of brassicaceae for optimum phytochemical composition. *J Agric Food Chem.* 2012;60(45):11409-11420. doi:10.1021/jf302863c.
- 72. Choi SH, Ryu DK, Park SY, Ann KG, Lim YP, An GH. Composition analysis between kohlrabi (*Brassica oleracea* var. *gongylodes*) and radish (*Raphanus sativus*). Kor J Hort Sci Technol. 2010;28:469-475.
- 73. Sharma I, Aaradhya M, Kodikonda M, Naik PR. Antihyperglycemic, antihyperlipidemic and antioxidant activity of phenolic rich extract of Brassica oleraceae var gongylodes on streptozotocin induced Wistar rats. *Springerplus*. 2015;4:212. doi:10.1186/s40064-015-0948-0.
- 74. Miranda-Osorio PH, Castell-Rodríguez AE, Vargas-Mancilla J, et al. Protective Action of Carica papaya on β-Cells in Streptozotocin-Induced Diabetic Rats. *Int J Environ Res Public Health*. 2016;13(5):446. doi:10.3390/ijerph13050446.
- 75. Juárez-Rojop IE, Díaz-Zagoya JC, Ble-Castillo JL, et al. Hypoglycemic effect of Carica papaya leaves in streptozotocin-induced diabetic rats. *BMC Complement Altern Med.* 2012;12:236. doi:10.1186/1472-6882-12-236
- 76. Raffaelli F, Nanetti L, Montecchiani G, et al. In vitro effects of fermented papaya (Carica papaya, L.) on platelets obtained from patients with type 2 diabetes. *Nutr Metab Cardiovasc Dis.* 2015;25(2):224-229. doi:10.1016/j.numecd.2014.10.013.
- 77. Sasidharan S, Sumathi V, Jegathambigai NR, Latha LY. Antihyperglycaemic effects of ethanol extracts of Carica papaya and Pandanus amaryfollius leaf in

streptozotocin-induced diabetic mice. *Nat Prod Res.* 2011;25(20):1982-1987. doi:10.1080/14786419.2010.523703.

- 78. Maniyar Y, Bhixavatimath P. Antihyperglycemic and hypolipidemic activities of aqueous extract of Carica papaya Linn. leaves in alloxan-induced diabetic rats. *J Ayurveda Integr Med*. 2012;3(2):70-74. doi:10.4103/0975-9476.96519.
- 79. Danese C, Esposito D, D'Alfonso V, Cirene M, Ambrosino M, Colotto M. Plasma glucose level decreases as collateral effect of fermented papaya preparation use. *Clin Ter.* 2006;157(3):195-198. PMID: 16900843.
- 80. Oboh G, Akinbola IA, Ademosun, AO, Sanni, DM, Odubanjo OV, Olasehinde TA, et al. Essential oil from clove bud (*Eugenia aromatic* Kuntze) inhibit key enzymes relevant to the management of Type-2 diabetes and some pro-oxidant induced lipid peroxidation in rats pancreas *in vitro*. *J. Oleo Sci.* 2015;64(7):775-782. doi: 10.5650/jos.ess14274.
- 81. Canal JR, Torres, MD, Romero A, Pérez, C. A chloroform extract obtained from a decoction of Ficus carica leaves improves the cholesterolaemic status of rats with streptozotocin-induced diabetes. *Acta Physiologica Hungarica*. 2000;87(1):71–76.
- 82. Arafa EA, Hassan W, Murtaza G, Buabeid MA. Ficus carica and Sizigium cumini regulate glucose and lipid parameters in high-fat diet and streptozocin-induced rats. *Journal of Diabetes Research Volume* 2020;6745873. doi: https://doi.org/10.1155/2020/6745873.
- 83. Irudayaraja SS, Christudasa S, Antonyb S, Duraipandiyanc V, Abdullahc AN, Ignacimuthua S. Protective effects of Ficus carica leaves on glucose and lipids levels, carbohydrate metabolism enzymes and bcells in type 2 diabetic rats. *Pharmaceutical biology*. 2017;55(1):1074–1081.
 - doi: 10.1080/13880209.2017.1279671.
- 84. Farsi E, Ahmad M, Hor SY, et al. Standardized extract of Ficus deltoidea stimulates insulin secretion and blocks hepatic glucose production by regulating the expression of glucose-metabolic genes

- in streptozitocin-induced diabetic rats. BMC Complement Altern Med. 2014;14:220. doi:10.1186/1472-6882-14-220.
- 85. Abdel-Rahman RF, Ezzat SM, Ogaly HA, et al. *Ficus deltoidea* extract downregulates protein tyrosine phosphatase 1B expression in a rat model of type 2 diabetes mellitus: a new insight into its antidiabetic mechanism. *J Nutr Sci.* 2020;9:e2. doi:10.1017/jns.2019.40.
- Kalman DS, Schwartz HI, Feldman S, Krieger DR. Efficacy and safety of Elaeis guineensis and Ficus deltoidea leaf extracts in adults with pre-diabetes. *Nutr J*. 2013;12:36. doi:10.1186/1475-2891-12-36.
- 87. Shrotri DS, Aiman R. The relationship of the post-absorptive state to the hypoglycemic action studies on Ficus bengalensis and Ficus glomerata. *Indian J Med Res.* 1960;48:162-168. PMID: 14446232.
- 88. Vasudevan K, Sophia D, Balakrishanan S, Manoharan S. Antihyperglycemic and antilipidperoxidative effects of Ficus racemosa (Linn.) bark extracts in alloxan induced diabetic rats. *J Med Sci.* 2017;7(3):330–338. doi: 10.3923/jms.2007.330.338.
- 89. Sophia D, Manoharan S. Hypolipidemic activities of Ficus racemosa Linn. bark in alloxan induced diabetic rats. *Afr J Tradit Complement Altern Med.* 2007;4(3):279-288. doi:10.4314/ajtcam.v4i3.31220.
- Patil KS, Warke PD, Chaturvedi SC. Hypoglycemic properties of Ficusglomerata fruits in alloxan-induced diabetic rats. *J Nat Remidies*. 6(2):120– 123.
- Wadood N, Nisar M, Rashid A, Wadood A, Gul-Nawab, Khan A. Effect of a compound recipe (medicinal plants) on serum insulin levels of alloxan induced diabetic rabbits. *J Ayub Med Coll Abbottabad*. 2007;19(1):32-38. PMID: 17867477.
- 92. Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. *J Ethnopharmacol*. 2003;84(1):105-108. doi:10.1016/s0378-8741(02)00144-7.

 Mandal SC, Mukharjee PK, Saha K, Das J, Pal M, Saha BP. Hypoglycemicactivity of Ficus racemosa L. (Moraceae) leaves in streptozotocin-induced diabeticrats. *Nat Prod Sci.* 1997;3(1):38–41.

- Patil VV, Pimprikar RB, Sutar NG, Barhate AL, Patil LS, Patil AP, et al. Antihyperglycemic activity of Ficus racemosa Linn leaves. *J Pharm Res.* 2009;(2):54–57.
- 95. Rahman NN, Khan M, Hasan R. Bioactive components from Ficus glomerata. *Pure Appl Chem.* 1994;66(10/11):2287–2290. doi: https://doi.org/10.1351/pac199466102 287.
- 96. Bwititi P, Musabayane CT. The effect of plant extracts on plasma glucose in rats. *Acta Med Biol.* 1997;45(4):167-169.
- 97. Musabayane CT, Gondwe M, Kamadyaapa DR, Chuturgoon AA, Ojewole JA. Effects of Ficus thonningii (Blume) Morarceae. stem-bark ethanolic extract on blood glucose, cardiovascular and kidney functions of rats, and on kidney cell lines of the proximal (LLC-PK1) and distal tubules (MDBK). Ren Fail. 2007;29(4):389-397. doi:10.1080/08860220701260735.
- 98. Minakawa M, Kawano A, Miura Y, Yagasaki K. Hypoglycemic effect of resveratrol in type 2 diabetic model db/db mice and its actions in cultured L6 myotubes and RIN-5F pancreatic β-cells. *J Clin Biochem Nutr.* 2011;48(3):237-244. doi:10.3164/jcbn.10-119.
- 99. Heim M, Johnson J, Boess F, et al. Phytanic acid, a natural peroxisome proliferator-activated receptor (PPAR) agonist, regulates glucose metabolism in rat primary hepatocytes. *FASEB J*. 2002;16(7):718-720. doi:10.1096/fj.01-0816fje.
- 100. Noor HS, Ismail NH, Kasim N, Mediani A, Zohdi RM, Ali AM et al. Urinary Metabolomics and Biochemical Analysis of Antihyperglycemic Effect of Ficus deltoidea Jack Varieties in Streptozotocin-Nicotinamide-Induced Diabetic Rats. *Appl Biochem Biotechnol*. 2020;192(1):1-21. doi: 10.1007/s12010-020-03304-y.
- 101. Nurdiana S, Goh YM, Ahmad H, et al. Changes in pancreatic histology, insulin secretion and oxidative status in diabetic rats following treatment with Ficus

- deltoidea and vitexin. *BMC Complement Altern Med*. 2017;17(1):290. doi:10.1186/s12906-017-1762-8.
- 102. Uzzaman R, Ghaffar M. Anti-diabetic and hypolipidemic effects of extract from the seed of Gossypium herbaceum L. in Alloxan-induced diabetic rabbits. *Pak J Pharm Sci.* 2017;30(1):75-86. PMID: 28603116.
- 103. Miaffo D, Ntchapda F, Kamgue OG, Mahamad AT, Kamanyi A. Glucoselowering potential of *Guiera senegalensis* roots in a diabetic rat model. *Avicenna J Phytomed*. 2020;10(6):653-663. PMID: 33299821.
- 104. Ibrahim MA, Islam MS. Butanol fraction of Khaya senegalensis root modulates β-cell function and ameliorates diabetes-related biochemical parameters in a type 2 diabetes rat model. *J Ethnopharmacol*. 2014;154(3):832-838. doi: 10.1016/j.jep.2014.05.011.
- 105. Ibrahim MA, Koorbanally NA, Islam MS. Antioxidative activity and inhibition of key enzymes linked to type-2 diabetes (α-glucosidase and α- amylase) by Khaya senegalensis. *Acta Pharm.* 2004;64(3):311-324. doi: 10.2478/acph-2014-0025.
- 106. Bothon FT, Debiton E, Avlessi F, Forestier C, Teulade JC, Sohounhloue DK. In vitro biological effects of two anti-diabetic medicinal plants used in Benin as folk medicine. *BMC Complement Altern Med*. 2013;13:51. doi:10.1186/1472-6882-13-51.
- 107. Arayne MS, Sultana N, Mirza AZ, Zuberi MH, Siddiqui FA. In vitro hypoglycemic activity of methanolic extract of some indigenous plants. *Pak J Pharm Sci*. 2007;20(4):268-273. PMID: 17604247.
- 108. Singh S, Verma N, Karwasra R, Kalra P, Kumar P, Gupta YK. Safety and efficacy of hydroalcoholic extract from *Lawsonia inermis*leaves on lipid profile in alloxaninduced diabetic rats. *Ayu*. 2015;36(1):107–112. doi: 10.4103/0974-8520.168999.
- 109. Ganogpichayagrai A, Palanuvej C, Ruangrungsi N. Antidiabetic and anticancer activities of *Mangifera indica* cv. Okrong leaves. *J Adv Pharm Technol Res.* 2017;8(1):19–24. doi: 10.4103/2231-4040.197371.

- 110. Ojo OA, Afon AA, Ojo AB, Ajiboye BO, Oyinloye BE, Kappo AB. Inhibitory effects of solvent-partitioned fractions of two Nigerian herbs (Spondias mombin Linn. and Mangifera indica L.) on α-Amylase and α- Glucosidase. *Antioxidants* (*Basel*).2018;7(6):73. doi: 10.3390/antiox7060073.
- 111. Villas Boas GR, Rodrigues Lemos JM, de Oliveira MW, et al. Aqueous extract from Mangifera indica Linn. (Anacardiaceae) leaves exerts long-term hypoglycemic effect, increases insulin sensitivity and plasma insulin levels on diabetic Wistar rats. *PLoS One*. 2020;15(1):e0227105. doi:10.1371/journal.pone.0227105.
- 112. Saleem M, Tanvir M, Akhtar MF, Iqbal M, Saleem A. Antidiabetic Potential of Mangifera indica L. cv. Anwar Ratol Leaves: Medicinal Application of Food Wastes. Medicina (Kaunas). 2019;55(7):353. doi:10.3390/medicina55070353.
- 113. Irondi EA, Oboh G, Akindahunsi AA. Antidiabetic effects of Mangifera indica Kernel Flour-supplemented diet in streptozotocin-induced type 2 diabetes in rats. *Food Sci Nutr.* 2016;4(6):828-839. doi: 10.1002/fsn3.348.
- 114. Villarruel-López A, López-de la Mora DA, Vázquez-Paulino OD, et al. Effect of Moringa oleifera consumption on diabetic rats. *BMC Complement Altern Med*. 2018;18(1):127. doi:10.1186/s12906-018-2180-2.
- 115. López M, Ríos-Silva M, Huerta M, et al. Effects of Moringa oleifera leaf powder on metabolic syndrome induced in male Wistar rats: a preliminary study. *J Int Med Res*. 2018;46(8):3327-3336. doi:10.1177/0300060518781726.
- 116. Paula PC, Sousa DO, Oliveira JT, et al. A Protein isolate from Moringa oleifera leaves has hypoglycemic and antioxidant effects in alloxan-induced diabetic mice. *Molecules*. 2017;22(2):271. doi:10.3390/molecules22020271.
- 117. Jaiswal D, Kumar Rai P, Kumar A, Mehta S, Watal G. Effect of Moringa oleifera Lam. leaves aqueous extract therapy on hyperglycemic rats. *J Ethnopharmacol*. 2009;123(3):392-396. doi:10.1016/j.jep.2009.03.036.

- 118. Yassa HD, Tohamy AF. Extract of Moringa oleifera leaves ameliorates streptozotocin-induced Diabetes mellitus in adult rats. *Acta Histochem*. 2014;116(5):844-854. doi:10.1016/j.acthis.2014.02.002.
- 119. Abd Eldaim MA, Shaban Abd Elrasoul A, Abd Elaziz SA. An aqueous extract from Moringa oleifera leaves ameliorates hepatotoxicity in alloxan-induced diabetic rats. *Biochem Cell Biol*. 2017;95(4):524-530. doi:10.1139/bcb-2016-0256.
- 120. Khan W, Parveen R, Chester K, Parveen S, Ahmad S. Hypoglycemic potential of aqueous extract of *Moringa oleifera* leaf and *In Vivo* GC-MS metabolomics. *Front Pharmacol*. 2017;8:577. doi:10.3389/fphar.2017.00577.
- 121. Olayaki LA, Irekpita JE, Yakubu MT, Ojo OO. Methanolic extract of Moringa oleifera leaves improves glucose tolerance, glycogen synthesis and lipid metabolism in alloxan-induced diabetic rats. *J Basic Clin Physiol Pharmacol*. 2015;26(6):585-593. doi:10.1515/jbcpp-2014-0129.
- 122. Omodanisi EI, Aboua YG, Chegou NN, Oguntibeju OO. Hepatoprotective, Antihyperlipidemic, and Antiinflammatory Activity of Moringa oleifera in Diabetic-induced Damage in Male Wistar Rats. Pharmacognosy Res. 2017;9(2):182-187. doi:10.4103/0974-8490.204651.
- 123. Alejandra Sánchez-Muñoz M, Valdez-Solana MA, Campos-Almazán MI, et al. Streptozotocin-induced adaptive modification of mitochondrial supercomplexes in liver of wistar rats and the protective effect of *Moringa oleifera* lam. *Biochem Res Int.* 2018;2018:5681081. doi:10.1155/2018/5681081.
- 124. Olurishe C, Kwanashie H, Zezi A, Danjuma N, Mohammed B. Chronic administration of ethanol leaf extract of Moringa oleifera Lam. (Moringaceae) may compromise glycaemic efficacy of Sitagliptin with no significant effect in retinopathy in a diabetic rat model. *J Ethnopharmacol*. 2016;194:895-903. doi:10.1016/j.jep.2016.10.065.
- 125. Tang Y, Choi EJ, Han WC, et al. Moringa oleifera from cambodia ameliorates oxidative stress, hyperglycemia, and

kidney dysfunction in Type 2 diabetic mice. *J Med Food*. 2017;20(5):502-510. doi:10.1089/jmf.2016.3792.

- 126. Al-Malki AL, El Rabey HA. The antidiabetic effect of low doses of Moringa oleifera Lam. seeds on streptozotocin induced diabetes and diabetic nephropathy in male rats. *Biomed Res Int*. 2015;2015:381040. doi:10.1155/2015/381040.
- 127. Jaja-Chimedza A, Zhang L, Wolff K, et al. A dietary isothiocyanate-enriched moringa (*Moringa oleifera*) seed extract improves glucose tolerance in a high-fat-diet mouse model and modulates the gut microbiome. *J Funct Foods*. 2018;47:376-385. doi:10.1016/j.jff.2018.05.056.
- 128. Wang F, Zhong HH, Chen WK, et al. Potential hypoglycaemic activity phenolic glycosides from Moringa oleifera seeds. *Nat Prod Res.* 2017;31(16):1869-1874. doi:10.1080/14786419.2016.1263846.
- 129. Raafat K, Hdaib F. Neuroprotective effects of Moringa oleifera: Bio-guided GC-MS identification of active compounds in diabetic neuropathic pain model published online ahead of print, 2017 Dec 12.. *Chin J Integr Med.* 2017;10.1007/s11655-017-2758-4. doi:10.1007/s11655-017-2758-4.
- 130. Gupta R, Mathur M, Bajaj VK, et al. Evaluation of antidiabetic and antioxidant activity of Moringa oleifera in experimental diabetes. *J Diabetes*. 2012;4(2):164-171. doi:10.1111/j.1753-0407.2011.00173.x.
- 131. Ogunyinka BI, Oyinloye BE, Osunsanmi FO, Kolanisi U, Opoku AR, Kappo AP. Protein isolate from *Parkia biglobosa* seeds improves dyslipidaemia and cardiac oxidative stress in Streptozotocin-induced diabetic rats. *Antioxidants* (*Basel*). 2019;8(10):481. doi:10.3390/antiox8100481.
- 132. Ibrahim MA, Habila JD, Koorbanally NA, Islam MS. Butanol fraction of Parkia biglobosa (Jacq.) G. Don leaves enhance pancreatic β-cell functions, stimulates insulin secretion and ameliorates other type 2 diabetes-associated complications in rats. *J Ethnopharmacol*. 2016;183:103-111. doi:10.1016/j.jep.2016.02.018.
- 133. Sule O, Godwin J, Abdu AR. Preliminary study of hypoglycemic effect of locust

- bean (Parkia biglobosa) on wistar albino rat. *J. Sci. Res. Rep.* 2015;4:467-472. doi:10.9734/JSRR/2015/8044.
- 134. Akinloye O, Akinmoladun AC, Farombi EO. Modulatory effect of Psidium guajava linn and ocimum gratissimum Linn on lipid profile and selected biochemical indices in rabbits fed high cholesterol diet. *J. Complement. Integr. Med.* 2010;7. doi:10.2202/1553-3840.1336.
- 135. Freire JM, Abreu CM, Duarte SM, Borges AF, Ribeiro LA. Evaluation of the protective effect of guava fruits and leaves on oxidative stress. *Acta Sci. Biol. Sci.* 2014;36(1):35–40. doi: 10.4025/actascibiolsci.v36i1.19839.
- 136. Oh WK, Lee CH, Lee MS, et al. Antidiabetic effects of extracts from Psidium guajava. *J Ethnopharmacol*. 2005;96(3):411-415. doi:10.1016/j.jep.2004.09.041.
- 137. Bahrani AHM, Zaheri H, Soltani N, Kharazmi F. Effect of the administration of Psidium guava leaves on blood glucose, lipid profiles and sensitivity of the vascular mesenteric bed to Phenylephrine in streptozotocin-induced diabetic rats. *J. Diabetes Mellit.* 2012;(2):138–145. doi: 10.4236/jdm.2012.21023.
- 138. Shen SC, Cheng FC, Wu NJ. Effect of guava (Psidium guajava Linn.) leaf soluble solids on glucose metabolism in type 2 diabetic rats. *Phytother Res*. 2008;22(11):1458-1464. doi:10.1002/ptr.2476.
- 139. Ogueri CC, Elekwa I, Ude VC, Ugbogu AE. Effect of aqueous extract of guava (Psidium guajava) leaf on blood glucose and liver enzymes in alloxan induced diabetic rats. *Br. J. Pharm. Res.* 2014;(4)9:1079-1087. doi: 10.9734/BJPR/2014/7244.
- 140. Shakeera BM, Sujatha K, Sridharan G, Manikandan R. Antihyperglycemic and antihyperlipidemic potentials of Psidium guajava in alloxan-induced diabetic rats. Asian J. Pharm. Clin. Res. 2013;6:88–89.
- 141. Soman S, Rauf AA, Indira M, Rajamanickam C. Antioxidant and antiglycative potential of ethyl acetate fraction of Psidium guajava leaf extract in streptozotocin-induced diabetic rats. *Plant*

- *Foods Hum Nutr.* 2010;65(4):386-391. doi:10.1007/s11130-010-0198-9.
- 142. Musabayane CT, Bwititi PT, Ojewole JA. Effects of oral administration of some herbal extracts on food consumption and blood glucose levels in normal and streptozotocin-treated diabetic rats. *Methods Find Exp Clin Pharmacol*. 2006;28(4):223-228. doi:10.1358/mf.2006.28.4.990202.
- 143. Asante DB, Effah-Yeboah E, Barnes P, et al. Antidiabetic Effect of Young and Old Ethanolic Leaf Extracts of Vernonia amygdalina: A Comparative Study. *J Diabetes Res.* 2016;2016:8252741. doi:10.1155/2016/8252741.
- 144. Ong KW, Hsu A, Song L, Huang D, Tan BK. Polyphenols-rich Vernonia amygdalina shows anti-diabetic effects in

- streptozotocin-induced diabetic rats. *J Ethnopharmacol*. 2011;133(2):598-607. doi:10.1016/j.jep.2010.10.046.
- 145. Nwanjo HU. Efficacy of aqueous leaf extract of vernonia amygdalina on plasma lipoprotein and oxidative status in diabetic rat models. *Niger J Physiol Sci.* 2005;20(1-2):39-42. PMID: 17220925.
- 146. Ibrahim MA, Islam MS. Effects of butanol fraction of Ziziphus mucronata root ethanol extract on glucose homeostasis, serum insulin and other diabetes-related parameters in a murine model for type 2 diabetes. *Pharm Biol.* 2017;55(1):416-422. doi:10.1080/13880209.2016.1242632.

(received 24.03.2021, published online 29.03.2021)

(одержано 24.03.2021, опубліковано 29.03.2021)

Conflict of interest

The author declares no conflict of interest.

Information about the authors

Ayodeji Oluwatobi Ojetunde

Academic degree: Bachelor of technology (B.Tech) in physiology.

Affiliations: Ladoke Akintola University of Technology, Ogbomoso, Nigeria.

Email: aoojetunde@gmail.com Phone number: +2347039155456